

# Omics Bootcamp

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Sept 19, 2014  
WSBS Genomics



# Outline



1. Applications of DNA Sequencing
  - Basic Concepts
  - Applications to Autism Genetics
  
2. “Functional” Assays
  - RNA-seq
  - Methyl-seq
  - ChIP-seq
  - Single Cell Sequencing

# Milestones in DNA Sequencing



(TIGR/Celera, 1995-2001)

# Inside the NY Genome Center

Sequencing Capacity Exceeds 2 Pbp/year (18,000 genomes / year)

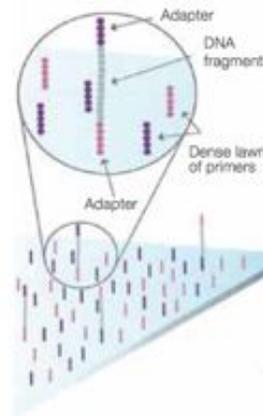


# Massively Parallel Sequencing

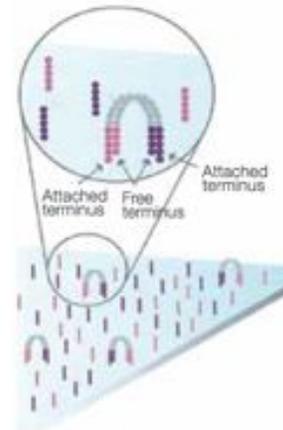


**Illumina HiSeq 2000**  
*Sequencing by Synthesis*

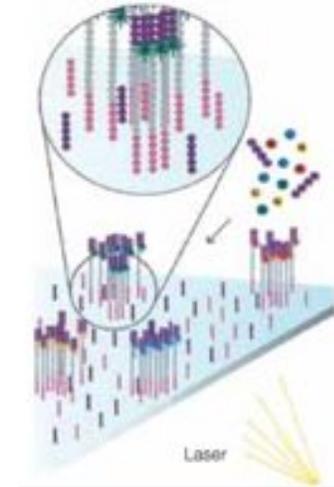
>60Gbp / day



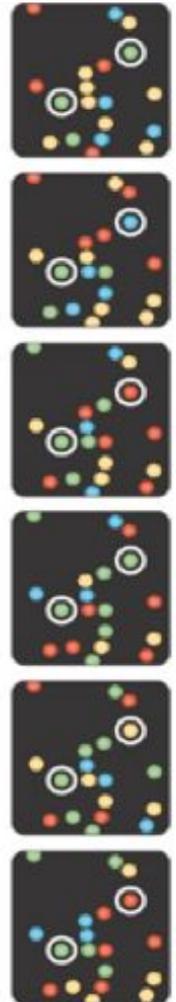
1. Attach



2. Amplify



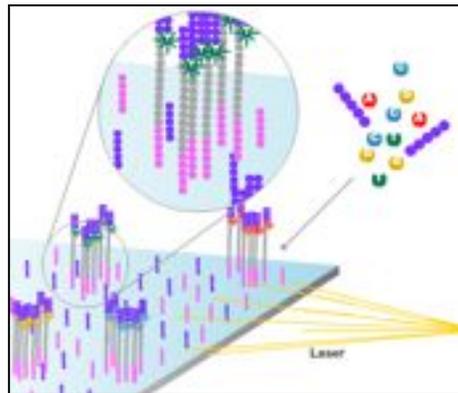
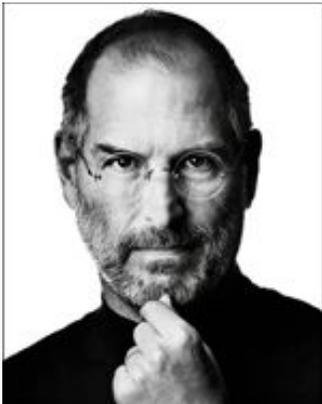
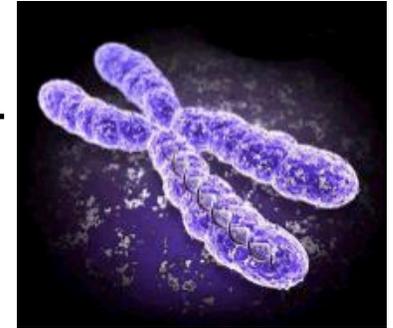
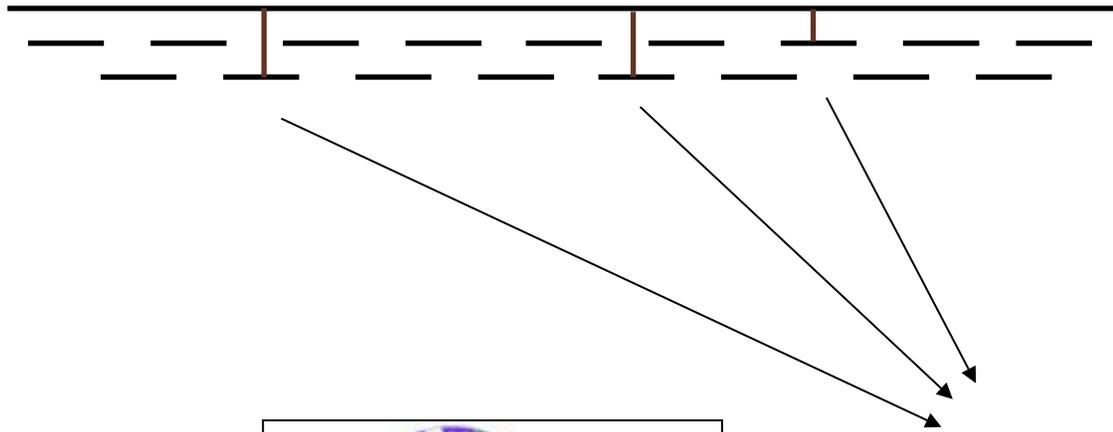
3. Image



Metzker (2010) Nature Reviews Genetics 11:31-46  
<http://www.youtube.com/watch?v=I99aKKHcxC4>

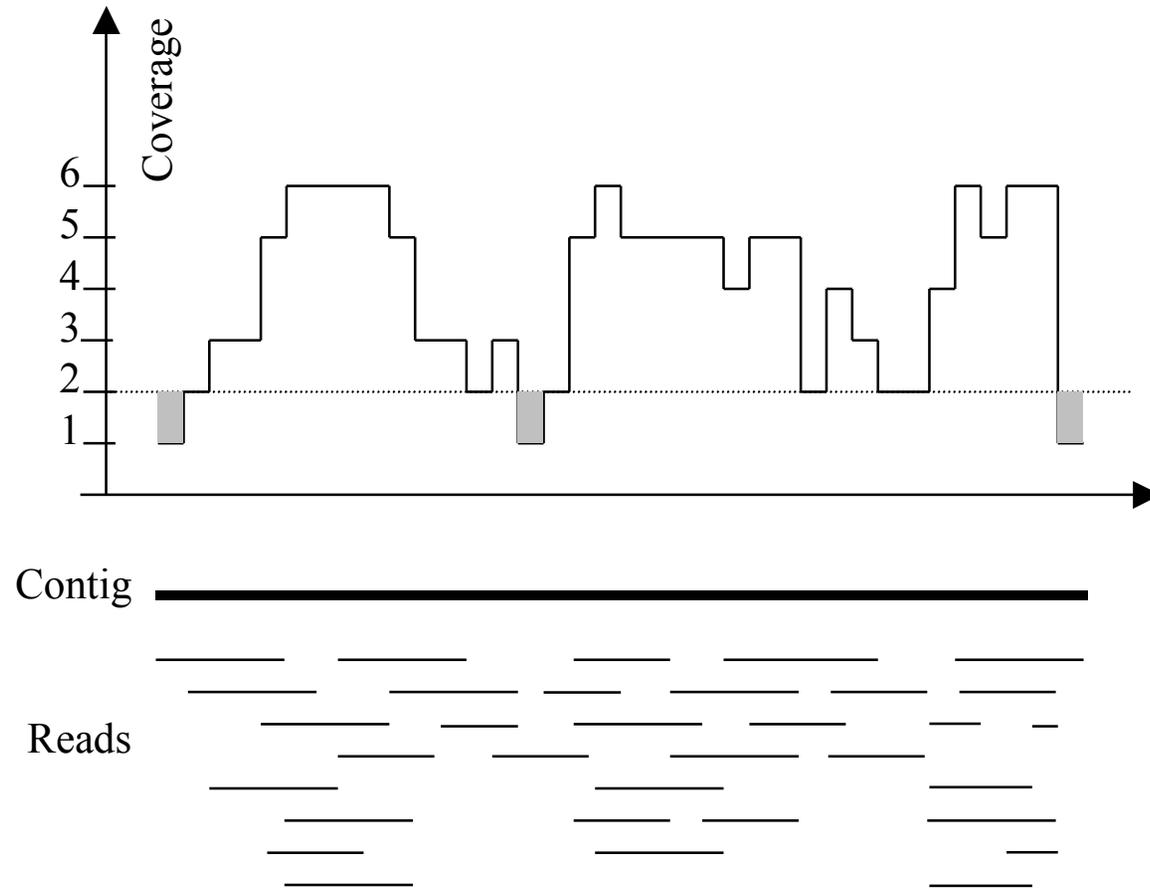
# Personal Genomics

How does your genome compare to the reference?



Heart Disease  
Cancer  
Creates magical  
technology

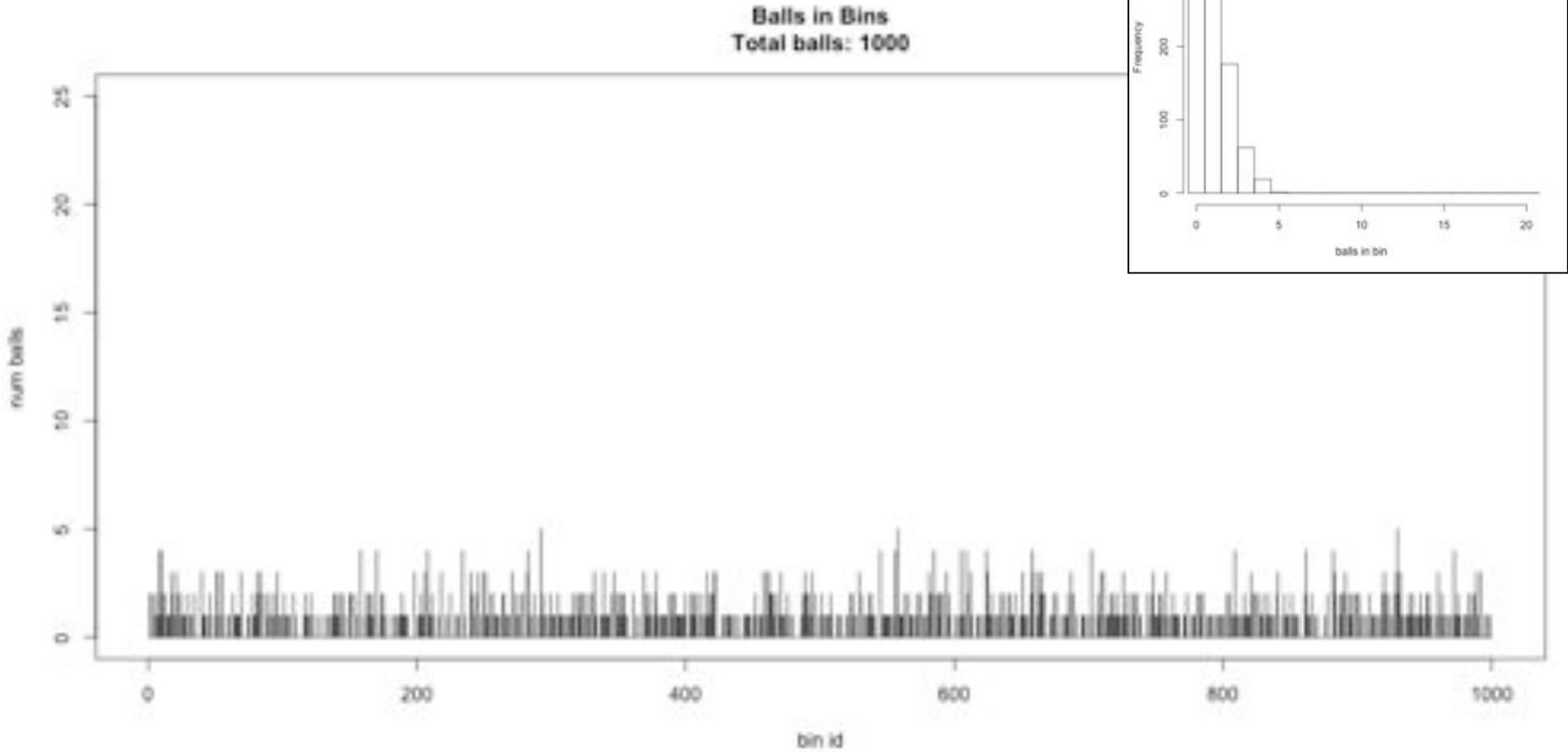
# Typical sequencing coverage



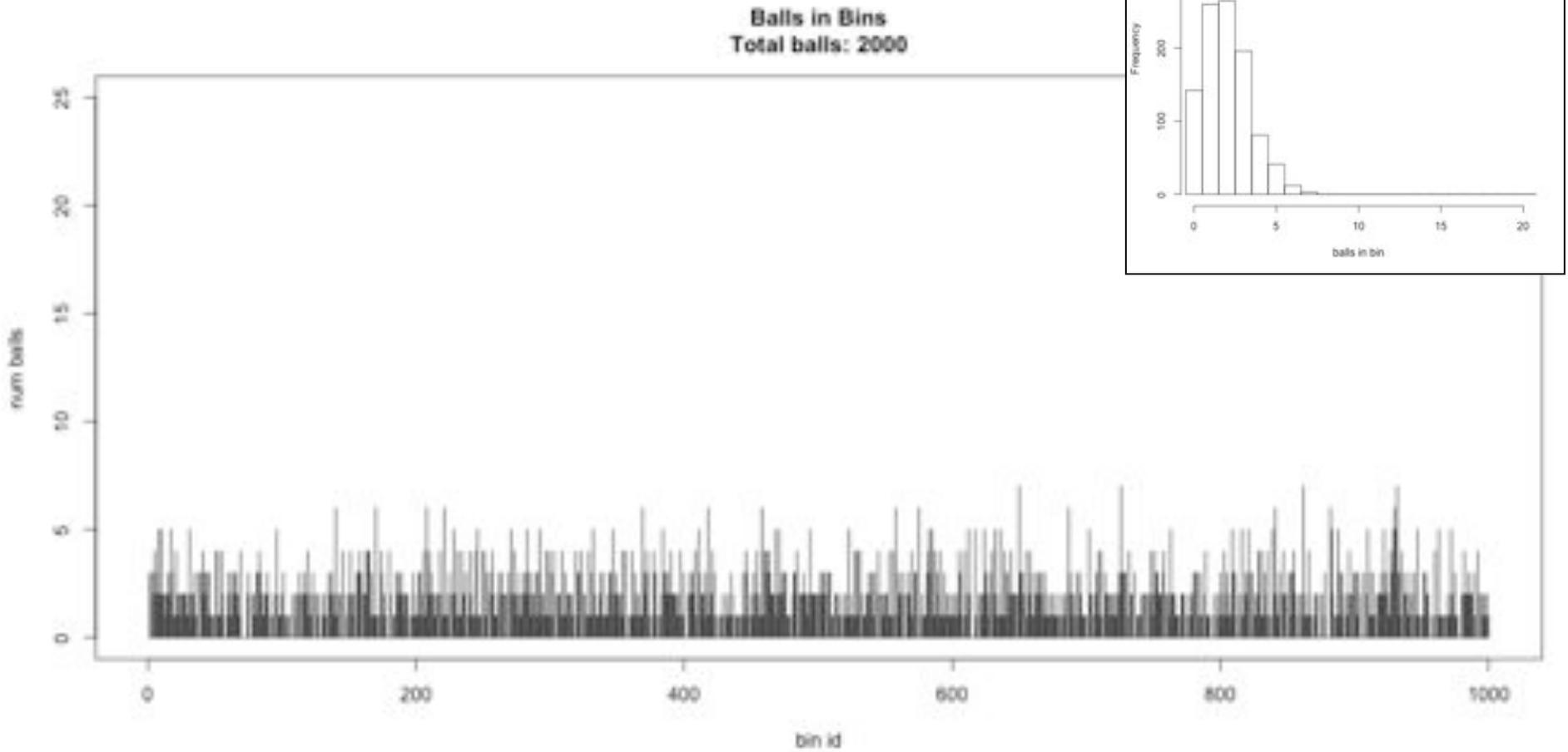
Imagine raindrops on a sidewalk

We want to cover the entire sidewalk but each drop costs \$1

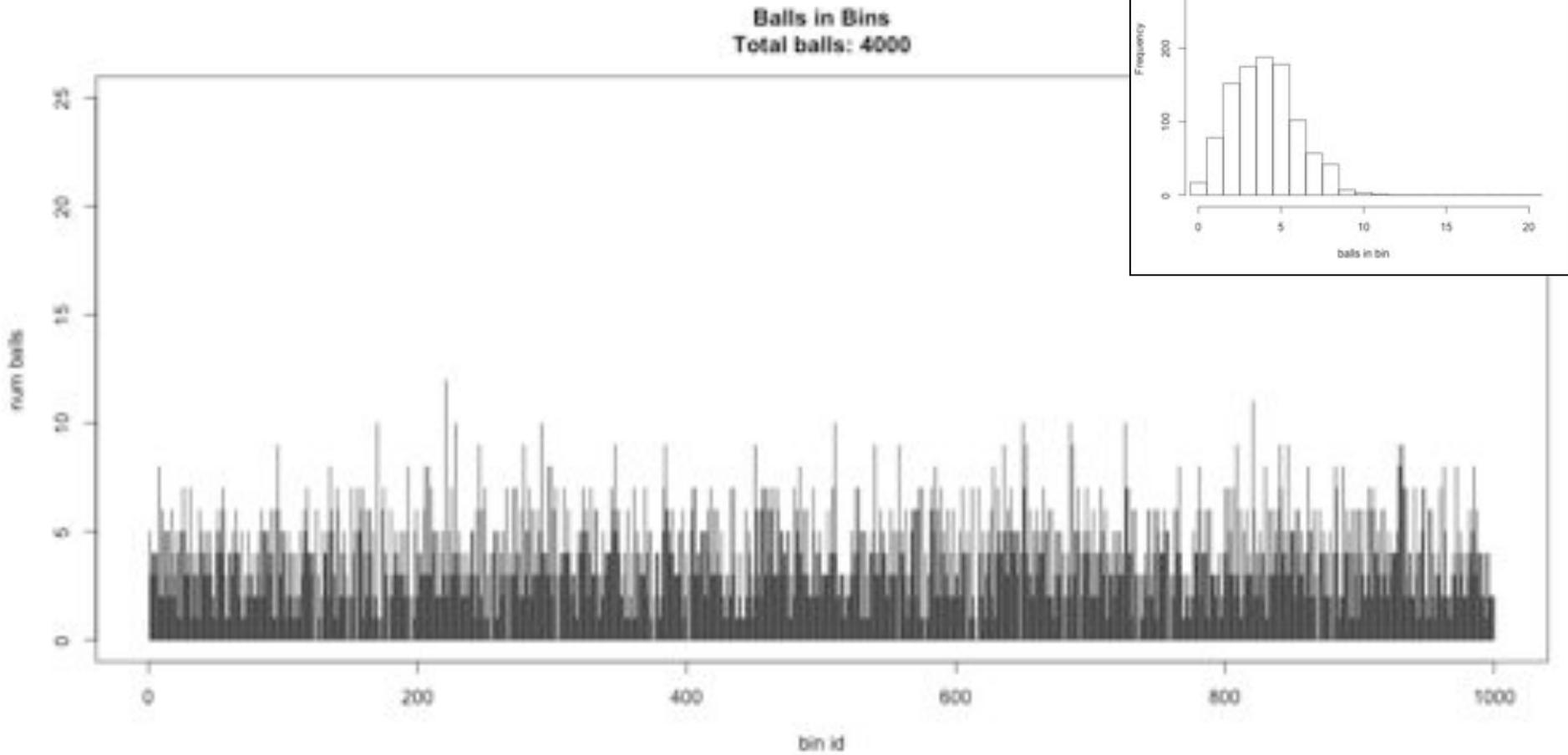
# Ix sequencing



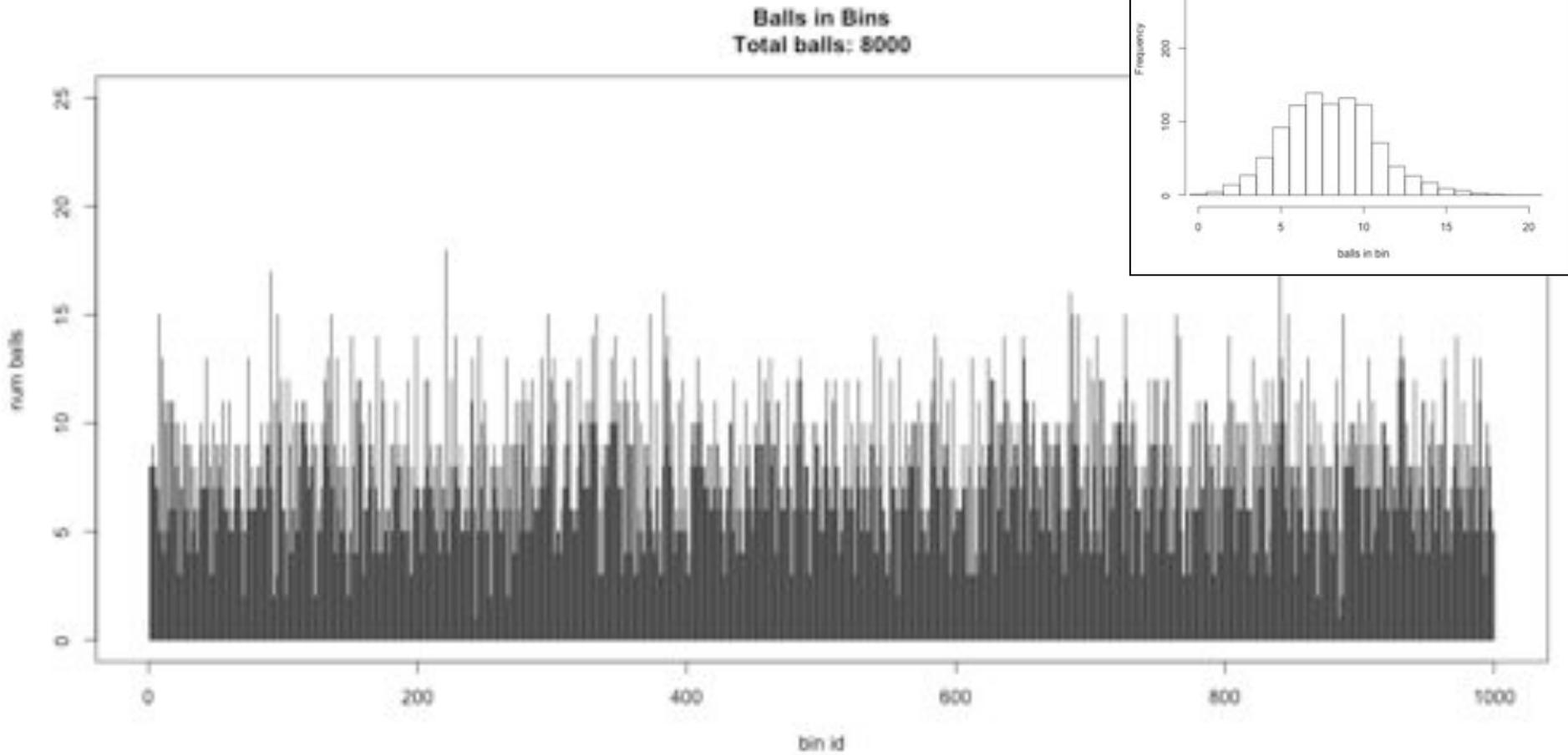
# 2x sequencing



# 4x sequencing



# 8x sequencing



# Poisson Distribution

The probability of a given number of events occurring in a fixed interval of time and/or space if these events occur with a known average rate and independently of the time since the last event.

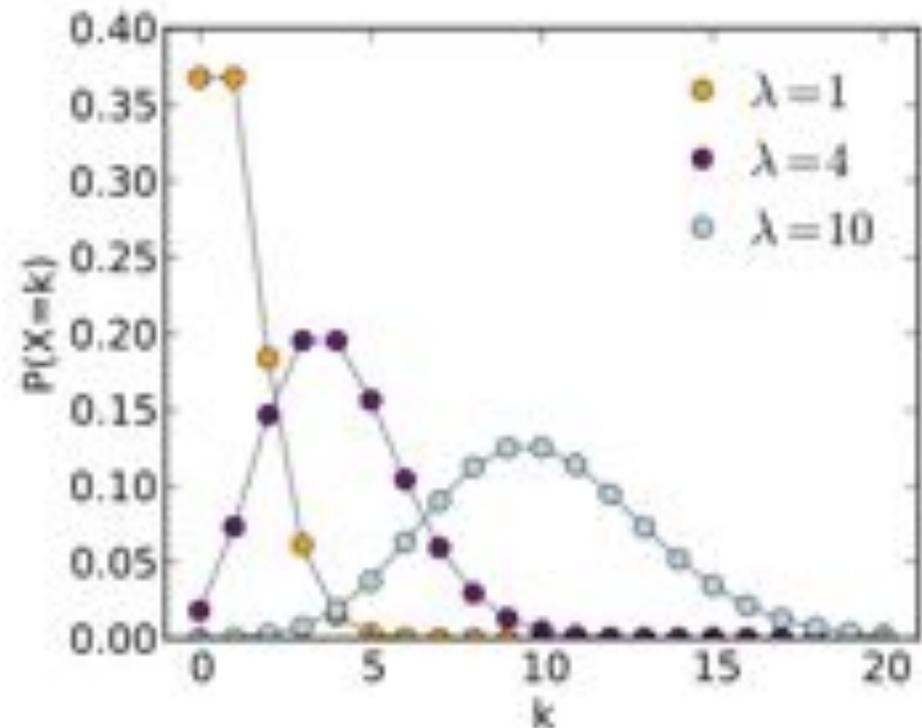
Formulation comes from the limit of the binomial equation

Resembles a normal distribution, but over the positive values, and with only a single parameter.

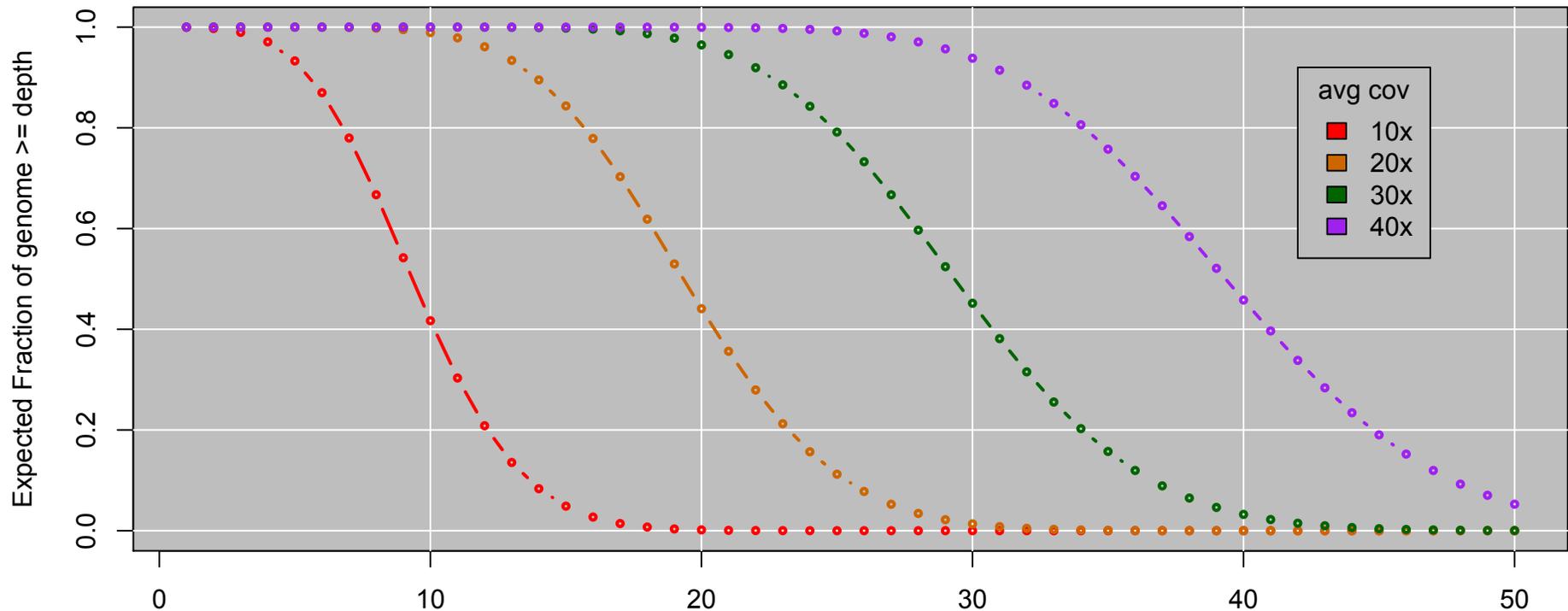
**Key property:**

- ***The standard deviation is the square root of the mean.***

$$P(k) = \frac{\lambda^k}{k!} e^{-\lambda}$$



# Genome Coverage Distribution



Expect Poisson distribution on depth

- Standard Deviation =  $\sqrt{\text{cov}}$

This is the mathematical model  $\Rightarrow$  reality may be much worse

- Double your coverage for diploid genomes
- Can use somewhat lower coverage in a population to find common variants

# Algorithms for Mapping & Genotyping

QB Week 1: Sept 29

### 1. Split read into segments

Read: CCAGTAGCTCTCAGCCTTATTTTACCCAGGCCTGTA  
 Read (reverse complement): TACAGGCCTGGGTAAAAAAGGCTGAGAGCTACTGG

Policy: extract 16 nt seed every 10 nt

Seeds:

```

    +, 0: CCAGTAGCTCTCAGCC      -, 0: TACAGGCCTGGGTAAA
    +, 10: TCAGCCTTATTTTACC     -, 10: GGTAAAAAAGGCTGA
    +, 20: TTTACCCAGGCCTGTA    -, 20: GGCTGAGAGCTACTGG
    
```

### 2. Lookup each segment and prioritize

Seeds:

```

    +, 0: CCAGTAGCTCTCAGCC
    +, 10: TCAGCCTTATTTTACC
    +, 20: TTTACCCAGGCCTGTA
    -, 0: TACAGGCCTGGGTAAA
    -, 10: GGTAAAAAAGGCTGA
    -, 20: GGCTGAGAGCTACTGG
    
```

Ungapped alignment with FM Index

Seed alignments (as B ranges):

```

    { [211, 212], [212, 214] }
    { [653, 654], [651, 653] }
    { [684, 685] }
    { }
    { }
    { [624, 625] }
    
```

### 3. Evaluate end-to-end match

Extension candidates:

```

    SA: 684, chr12:1955
    SA: 624, chr2:462
    SA: 211, chr4:762
    SA: 213, chr12:1935
    SA: 652, chr12:1945
    
```

SIMD dynamic programming aligner

SAM alignments:

```

    r1 0 chr12 1936 0
    36M * 0 0
    CCAGTAGCTCTCAGCCTTATTTTACCCAGGCCTGTA
    IHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHH
    AS:i:0 XS:i:-2 XN:i:0
    XM:i:0 XO:i:0 XG:i:0
    NM:i:0 MD:Z:36 YT:Z:UU
    YN:i:0
    ...
    
```

Heterozygous variant?      Homozygous variant

```

    ...CCATAG      GTGCGCCC      CGGAATT      GGTATAC...
    ...CCAT      CTATGTGCG      TCGGAATT      CGGTATAC
    ...CCAT GGCTATGTG      CTATCGGAAA      GCGGTATA
    ...CCA AGGCTAAT      CCTATCGGA      TTGCGGTA C...
    ...CCA AGGCTAAT      GCCCTATCG      TTTGCGGT C...
    ...CC AGGCTATA      GCCCTATCG      AAATTTGC      ATAC...
    ...CC TAGGCTATA      GCGCCCTA      AAATTTGC      GTATAC...
    ...CCATAGGCTAATGCGCCCTATCGCAATTTGCGGTATAC...
    
```

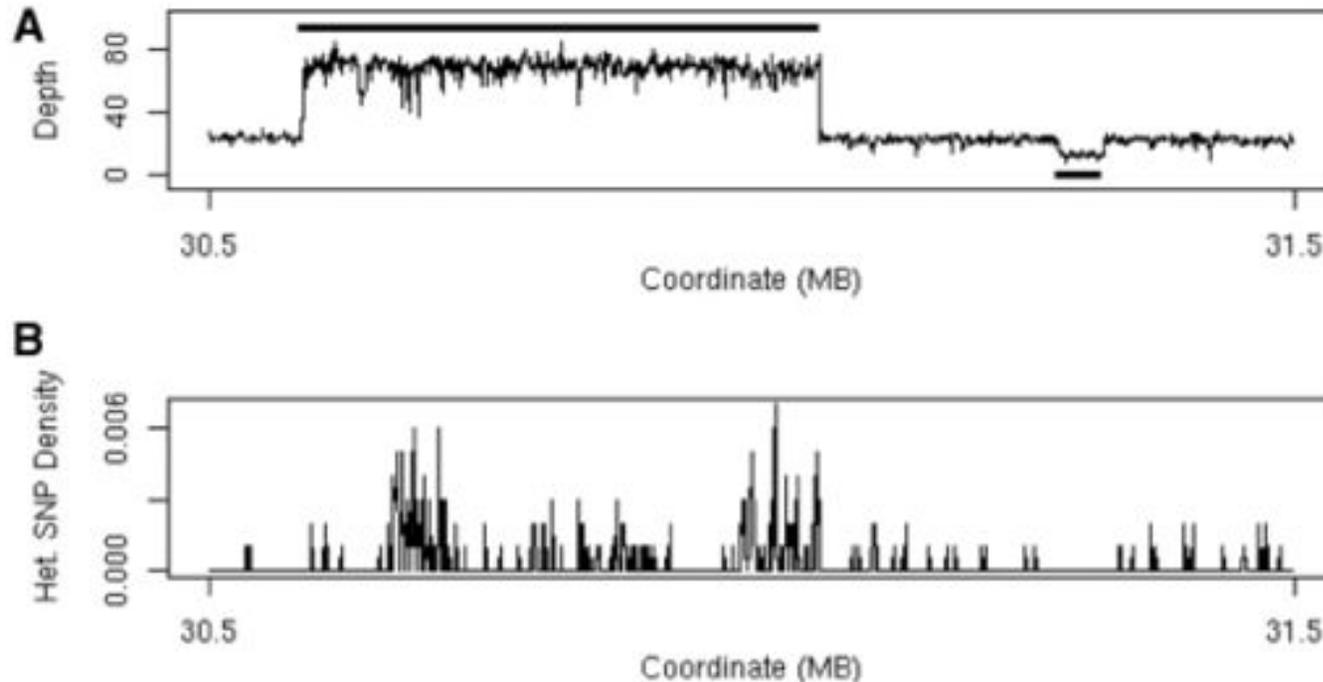
- Distinguishing SNPs from sequencing error typically a likelihood test of the coverage
  - Hardest to distinguish between errors and heterozygous SNP.
  - Coverage is the most important factor!
    - Target at least 10x, 30x more reliable

**Fast gapped-read alignment with Bowtie 2**  
 Langmead & Salzberg. (2012) *Nature Methods*. 9:357-359.

**The Sequence Alignment/Map format and SAMtools**  
 Li H et al. (2009) *Bioinformatics*. 25:16 2078-9

# CNV calling

*Beware of (Systematic) Errors*



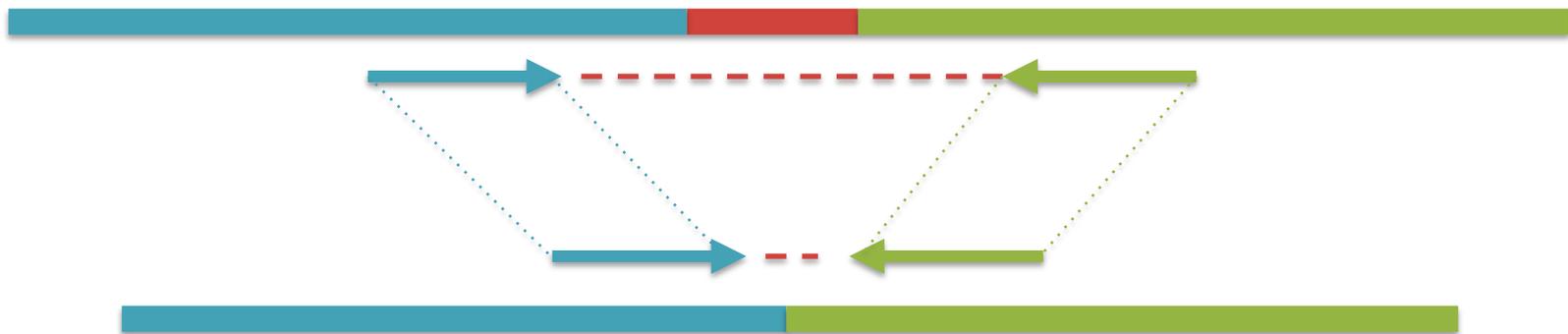
(A) Plot of sequencing depth across a one megabase region of A/J chromosome 17 clearly shows both a region of 3-fold increased copy number (30.6–31.1 Mb) and a region of decreased copy number (at 31.3 Mb).

Simpson J T et al. *Bioinformatics* 2010;26:565-567

- Identify CNVs through increased depth of coverage & increased heterozygosity
  - Segment coverage levels into discrete steps
  - Be careful of GC biases and mapping biases of repeats

# Structural Variations

Sample Separation: 2kbp



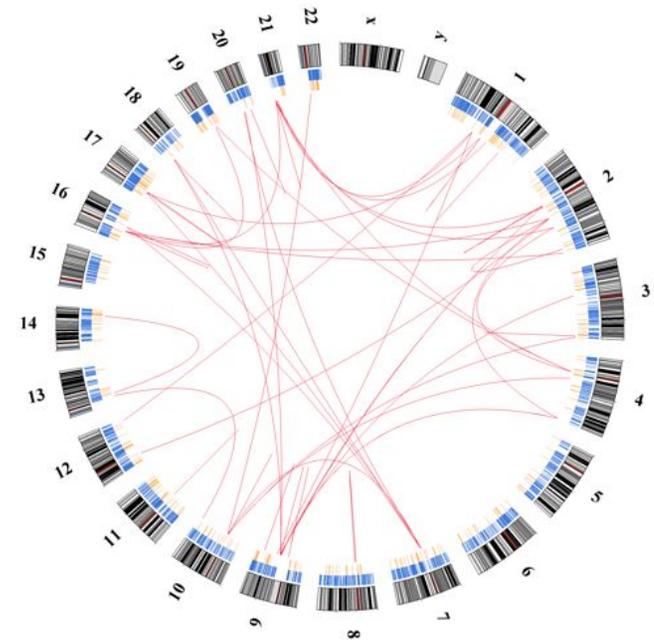
Mapped Separation: 1kbp

SVs tend to be flanked by repeats, making it hard to localize

- Cannot trust results from a single compress/expanded mate, look for a cluster of them
- Longer reads are the key to resolving them

Circos plot of high confidence SVs specific to esophageal cancer sample

- Red: SV links
- Orange: 375 cancer genes
- Blue: 4950 disease genes

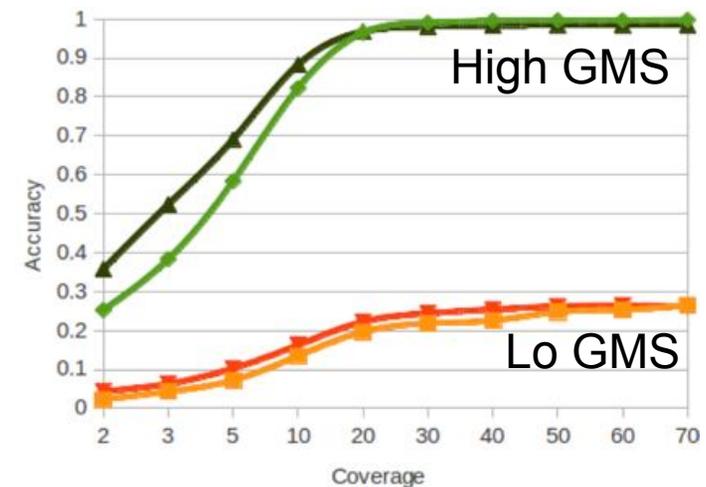


# Beware of Mapping Errors



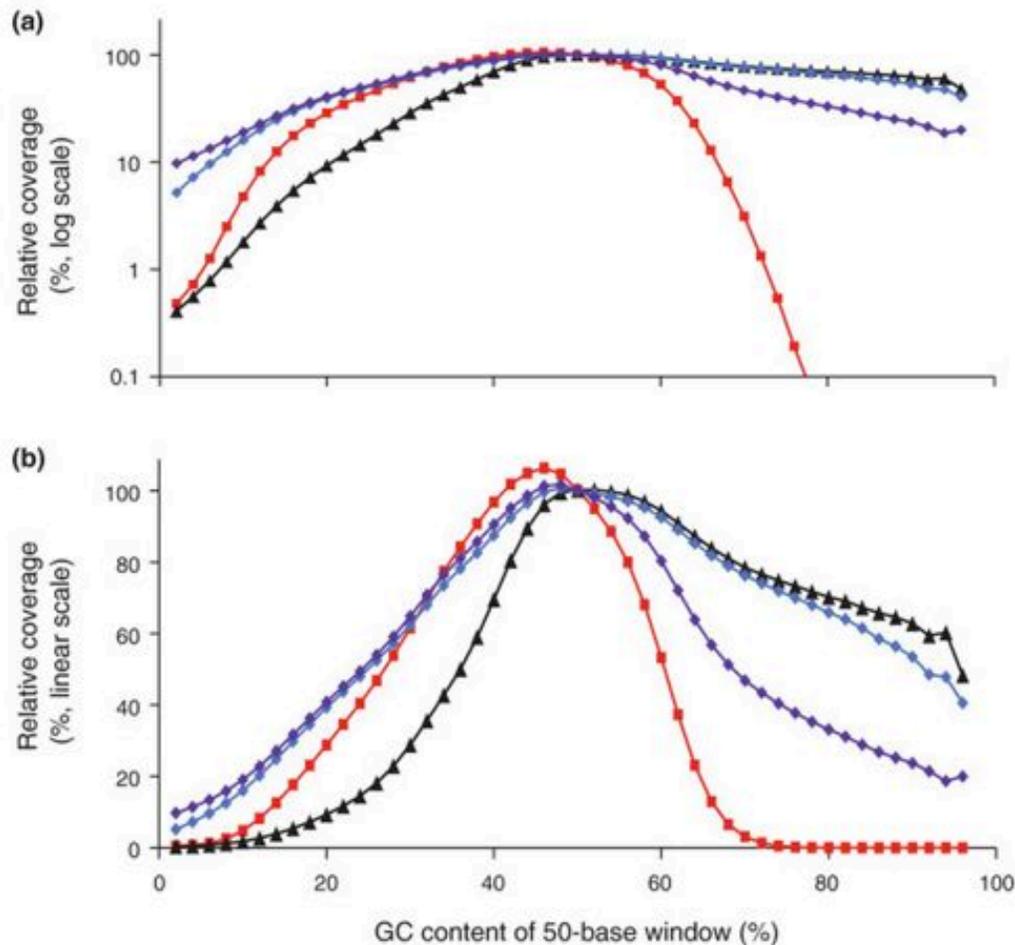
- Short read mapping is an essential for identifying mutations in the genome
  - Not every base of the genome can be mapped equally well, especially because of repeats
- Introduced a new probabilistic metric - the Genome Mappability Score - that quantifies how reliably reads can be mapped to every position in the genome
  - We have little power to measure 11-13% of the human genome, including of known clinically relevant variations
  - Errors in variation discovery are dominated by errors in low GMS regions

Species (build)	size	paired/single	whole (%)	transcription (%)
yeast (sc2)	12 Mbp	paired	94.85	95.04
		single	94.25	94.62
fly (dm3)	130 Mbp	paired	90.52	96.14
		single	89.70	95.94
mouse (mm9)	2.7 Gbp	paired	89.39	96.03
		single	87.47	94.75
human (hg19)	3.0 Gbp	paired	89.02	97.40
		single	87.79	96.38



**Genomic Dark Matter: The reliability of short read mapping illustrated by the GMS.**  
Lee and Schatz (2012) *Bioinformatics*. doi: 10.1093/bioinformatics/bts330

# Beware of GC Biases



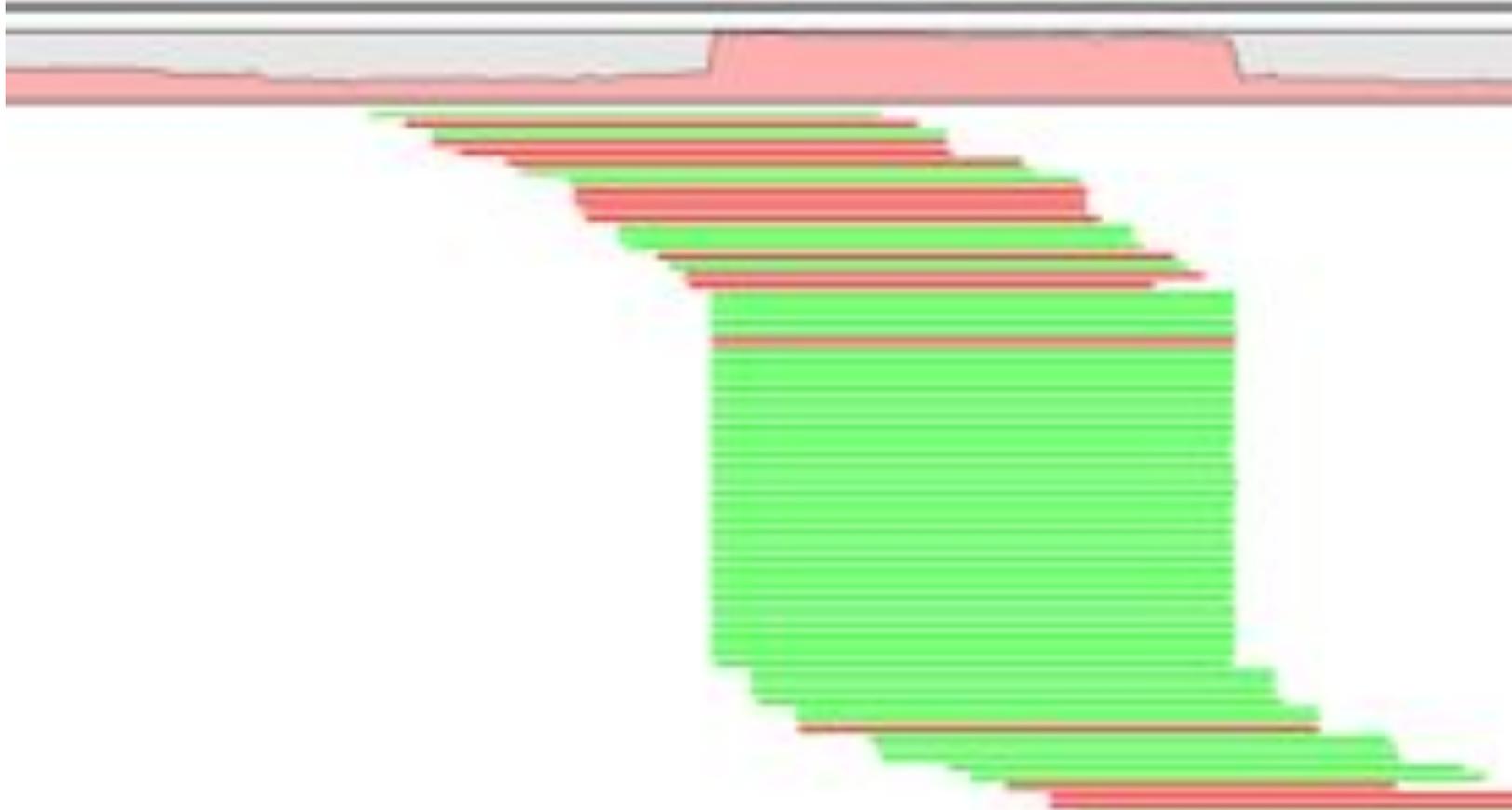
**Illumina sequencing does not produce uniform coverage over the genome**

- Coverage of extremely high or extremely low GC content will have reduced coverage in Illumina sequencing
- Biases primarily introduced during PCR; lower temperatures, slower heating, and fewer rounds minimize biases
- This makes it very difficult to identify variants (SNPs, CNVs, etc) in certain regions of the genome

**Analyzing and minimizing PCR amplification bias in Illumina sequencing libraries.**

Aird et al. (2011) *Genome Biology*. 12:R18.

# Beware of Duplicate Reads



**The Sequence alignment/map (SAM) format and SAMtools.**

Li et al. (2009) *Bioinformatics*. 25:2078-9

Picard: <http://picard.sourceforge.net>



# Genetic Basis of Autism Spectrum Disorders



## ***Complex disorders of brain development***

- Characterized by difficulties in social interaction, verbal and nonverbal communication and repetitive behaviors.
- Have their roots in very early brain development, and the most obvious signs of autism and symptoms of autism tend to emerge between 2 and 3 years of age.

## ***U.S. CDC identify around 1 in 68 American children as on the autism spectrum***

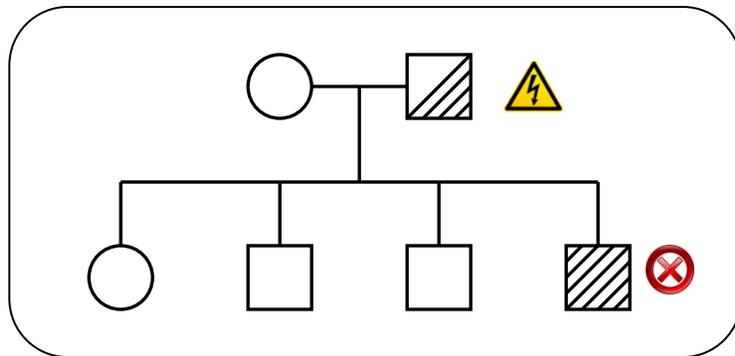
- Ten-fold increase in prevalence in 40 years, only partly explained by improved diagnosis and awareness.
- Studies also show that autism is four to five times more common among boys than girls.
- Specific causes remain elusive

## **What is Autism?**

<http://www.autismspeaks.org/what-autism>

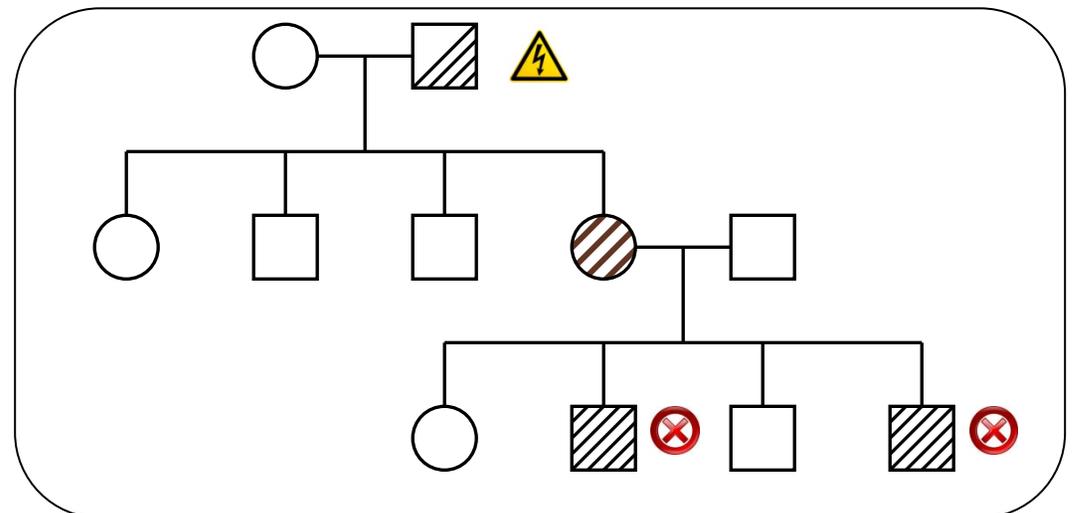
# Unified Model of Autism

## Sporadic Autism: 1 in 100



**Prediction:** De novo mutations of high penetrance contributes to autism, especially in low risk families with no history of autism.

## Familial Autism: 90% concordance in twins



### Legend



Sporadic mutation



Fails to procreate

**A unified genetic theory for sporadic and inherited autism**

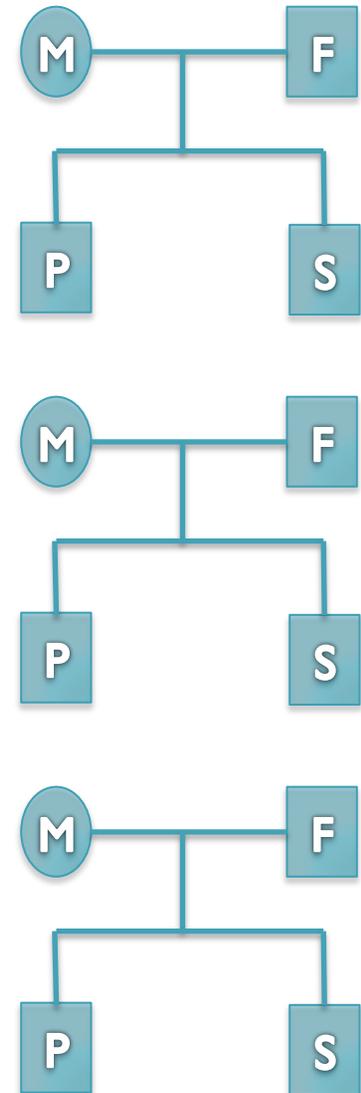
Zhao *et al.* (2007) *PNAS*. 104(31)12831-12836.

# Searching for the genetics behind human disorders and plant phenotypes

## ***Search Strategy***

- Currently uses whole exome short read resequencing for economic reasons
- Collaborate with Lyon, McCombie, Tuveson, and Wigler labs to examine the genetic basis of cancer, ASD, and other psychiatric disorders
- Also collaborating with the Lippman, Ware, and Gingeras labs to study high value crops

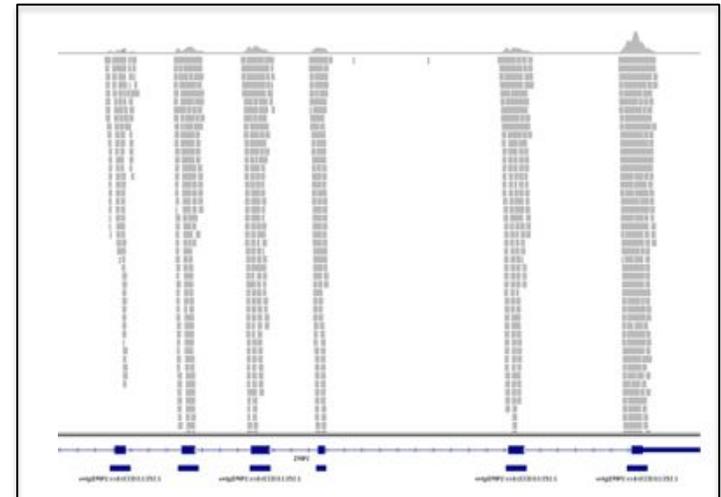
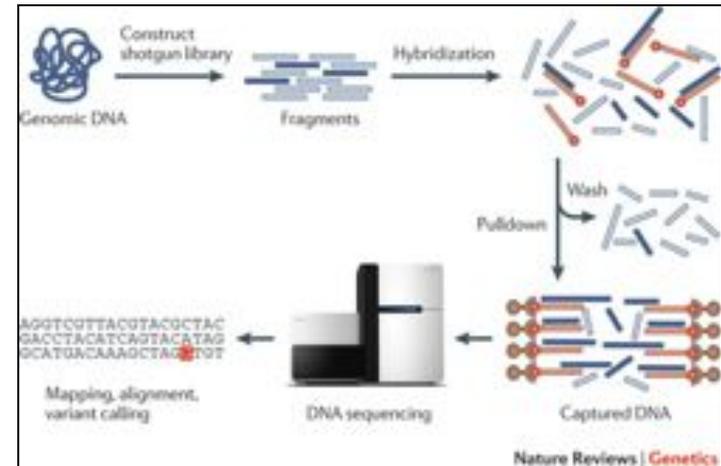
***Are there any genetic variants present in affected individuals, that are not present or are present at a substantially reduced rate in their relatives?***



# Exome-Capture Sequencing

## Exome-capture reduces the costs of sequencing

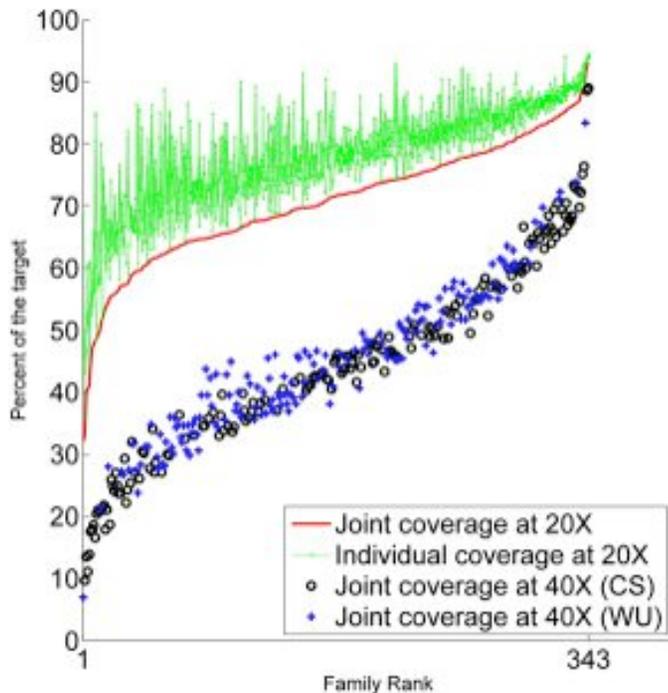
- Currently targets around 50Mbp of sequence: all exons plus flanking regions
- WGS currently costs ~\$2000 per sample, while WES currently costs ~\$400 per sample
- Coverage is highly localized around genes, although will get sparse coverage throughout rest of genome



## Exome sequencing as a tool for Mendelian disease gene discovery

Bamshad et al. (2011) *Nature Reviews Genetics*. 12, 745-755

# Exome sequencing of the SSC



## ***The year 2012 was an exciting year for autism genetics***

- 3 reports of >593 families from the Simons Simplex Collection (parents plus one child with autism and one non-autistic sibling)
- All attempted to find mutations enriched in the autistic children
- ***All used poor or no tools for indels:***
  - lossifov (343 families) and O’Roak (50 families) used GATK UnifiedGenotype
  - Sanders (200 families) didn’t attempt

## **De novo gene disruptions in children on the autism spectrum**

lossifov *et al.* (2012) *Neuron*. 74:2 285-299

## **De novo mutations revealed by whole-exome sequencing are strongly associated with autism**

Sanders *et al.* (2012) *Nature*. 485, 237–241.

## **Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations**

O’Roak *et al.* (2012) *Nature*. 485, 246–250.



# Scalpel: Haplotype Microassembly

DNA sequence **micro-assembly** pipeline for accurate detection and validation of *de novo* mutations (SNPs, indels) within exome-capture data.



## Features

1. Combine **mapping** and **assembly**
2. Exhaustive search of **haplotypes**
3. **De novo mutations**

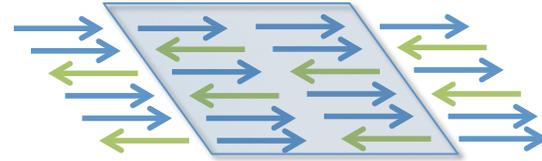


NRXN1 *de novo* SNP  
(auSSC12501 chr2:50724605)

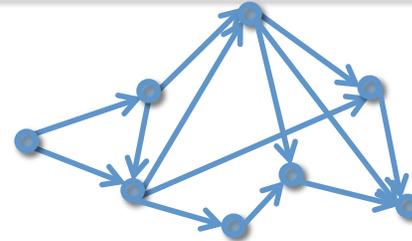
**Accurate de novo and transmitted indel detection in exome-capture data using microassembly.**  
Narzisi et al. (2014) *Nature Methods*. doi:10.1038/nmeth.3069

# Scalpel Algorithm

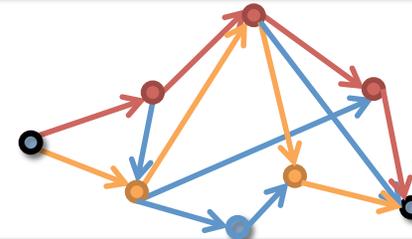
Extract reads mapping within the exon including (1) well-mapped reads, (2) soft-clipped reads, and (3) anchored pairs



Decompose reads into overlapping  $k$ -mers and construct de Bruijn graph from the reads



Find end-to-end haplotype paths spanning the region



Align assembled sequences to reference to detect mutations



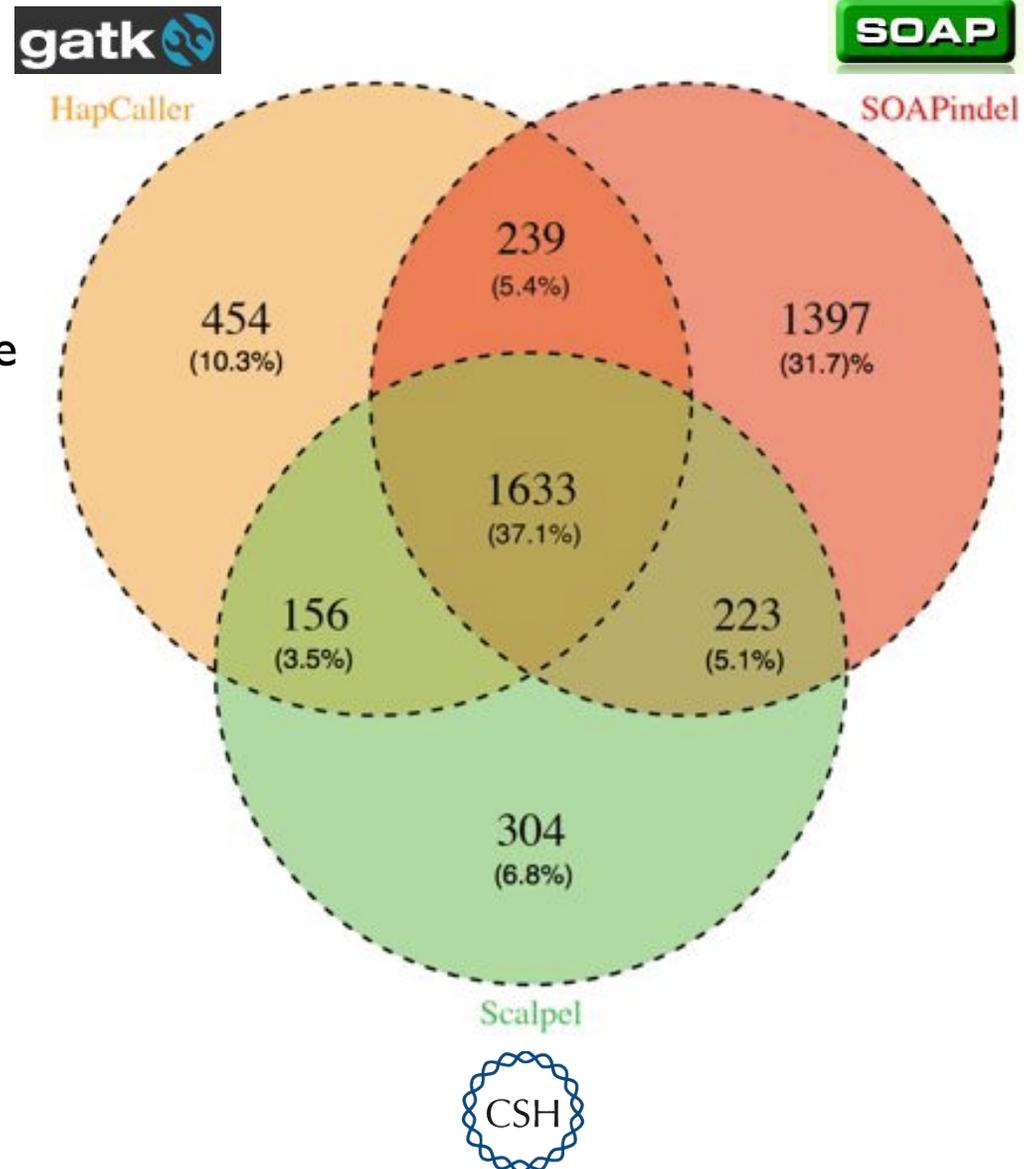
# Experimental Analysis & Validation

Selected one deep coverage exome for deep analysis

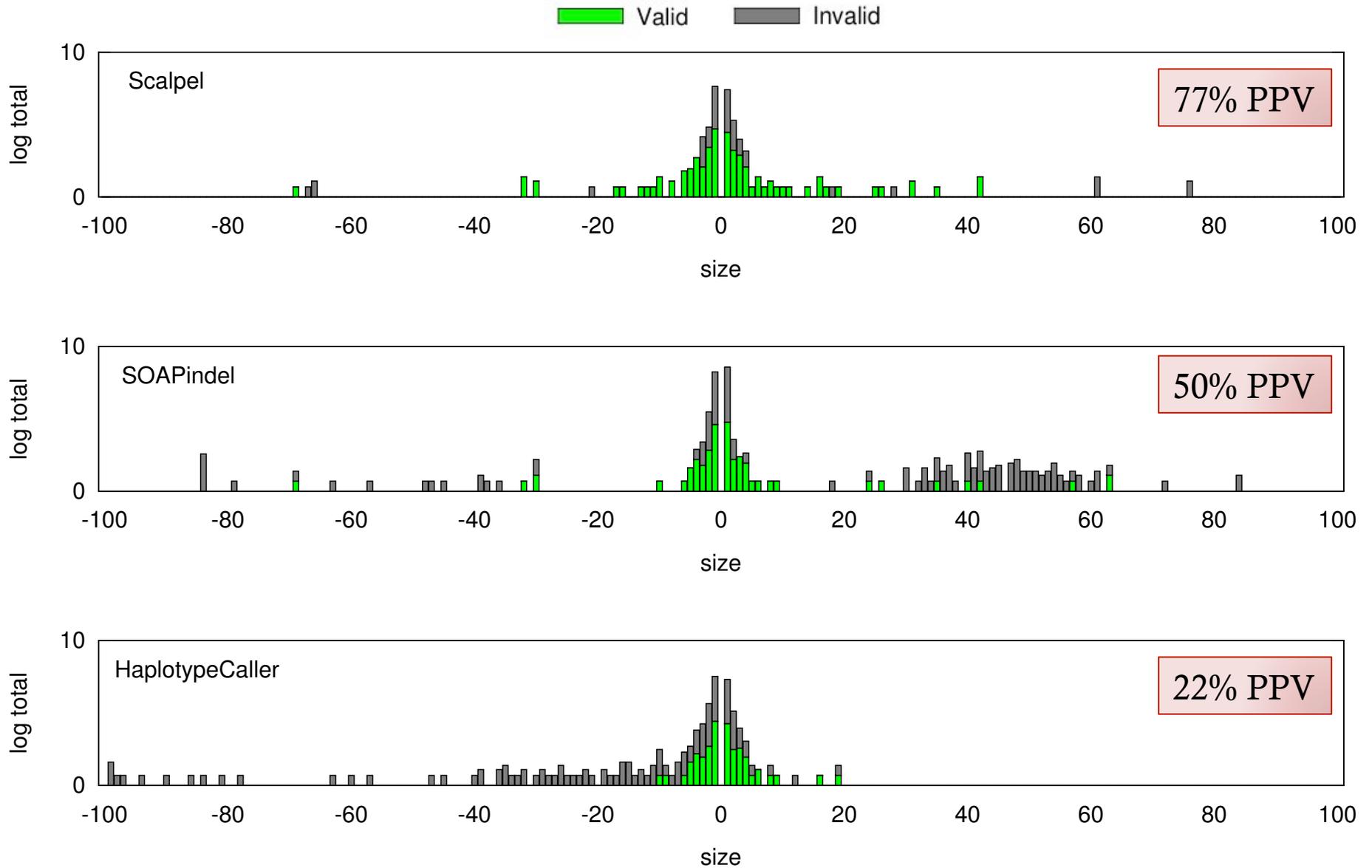
- Individual was diagnosed with ADHD and turrets syndrome
- 80% of the target at >20x coverage
- Evaluated with Scalpel, SOAPindiel, and GATK Haplotype Caller

1000 indels selected for validation

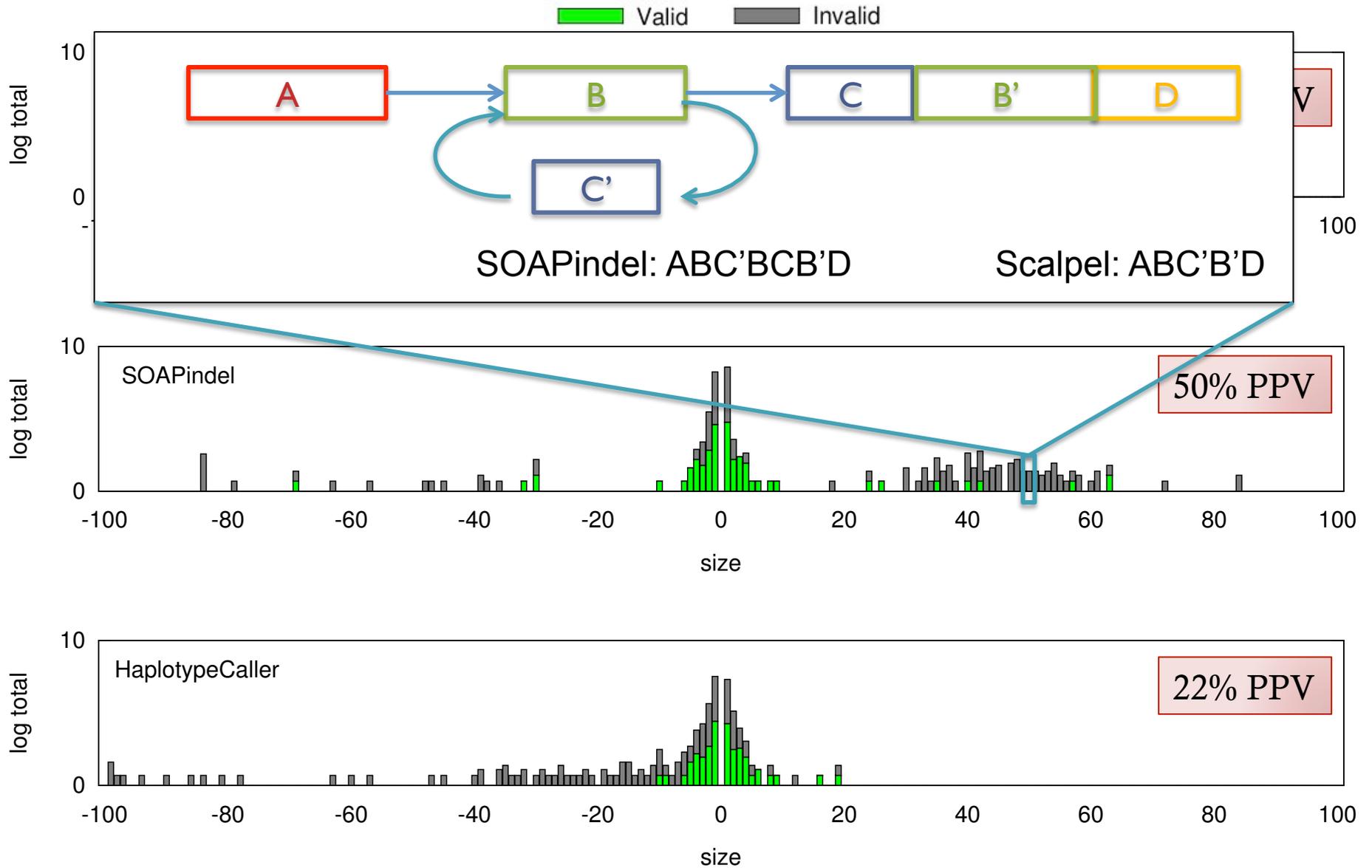
- 200 Scalpel
- 200 GATK Haplotype Caller
- 200 SOAPindiel
- 200 within the intersection
- 200 long indels (>30bp)



# Scalpel Indel Validation

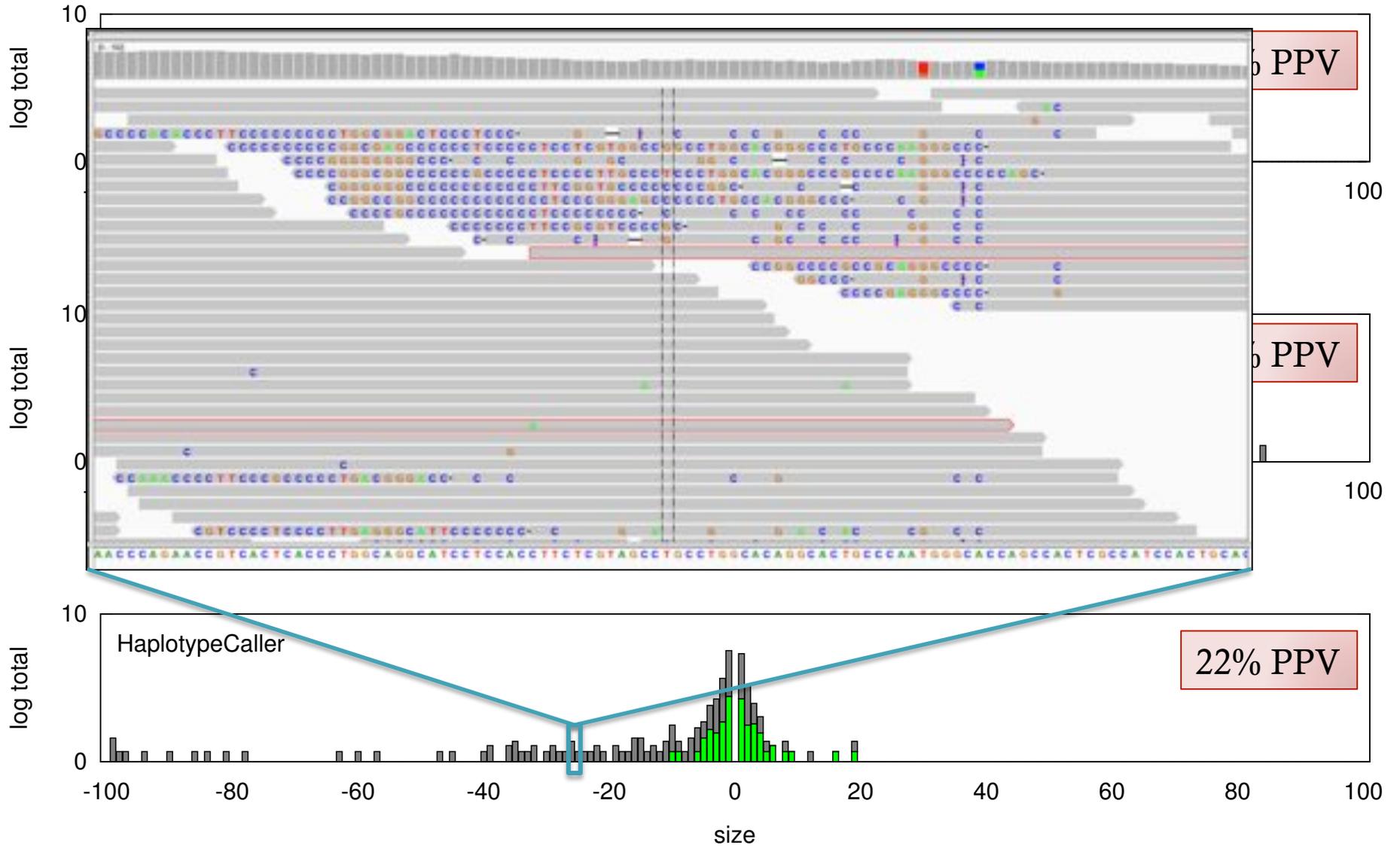


# Scalpel Indel Validation



# Scalpel Indel Validation

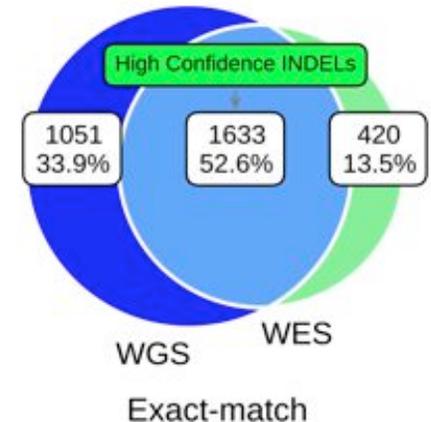
Valid Invalid



# Refined indel analysis

## Examine sources of indel errors

- Experimental validation of indels called from 30x whole genome vs. 110x whole exome of the same sample
- Most of the errors due to short microsatellite errors introduced during exome capture, also misses most long indels
- Recommend WGS for indel analysis instead

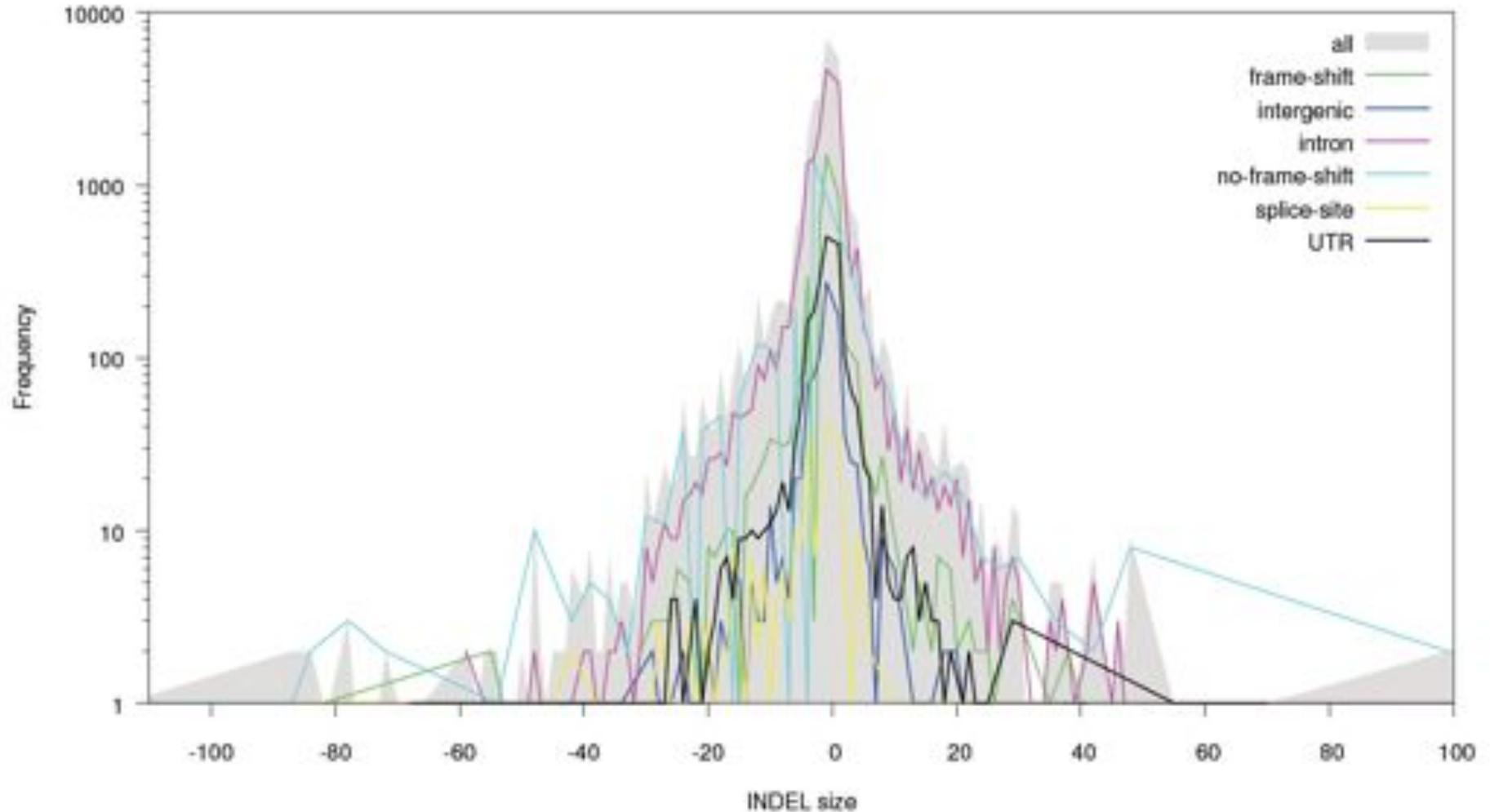


	All INDELS	Valid	PPV	INDELS >5bp	Valid (>5bp)	PPV (>5bp)
Intersection	160	152	95.0%	18	18	100%
WGS	145	122	84.1%	33	25	75.8%
WES	161	91	56.5%	1	1	100%

## Reducing INDEL calling errors in whole-genome and exome sequencing data

Fang, H, Wu, Y, Narzisi, G, O'Rawe, JA, Jimenez Barrón LT, Rosenbaum, J, Ronemus, M, Lossifov I, Schatz, MC<sup>§</sup>, Lyon, GL<sup>§</sup>  
<http://www.biorxiv.org/content/early/2014/06/10/006148>

# Revised Analysis of the SSC

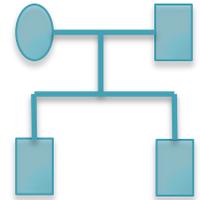


Constructed database of  $>1M$  transmitted and de novo indels  
Many new gene candidates identified, population analysis underway

# De novo mutation discovery and validation

## De novo mutations:

Sequences not inherited from your parents.



**Reference:** . . . **TCAAATCCTTTTAATAAAGAAGAGCTGACA** . . .

Father(1): . . . TCAAATCCTTTTAATAAAGAAGAGCTGACA . . .

Father(2): . . . TCAAATCCTTTTAATAAAGAAGAGCTGACA . . .

Mother(1): . . . TCAAATCCTTTTAATAAAGAAGAGCTGACA . . .

Mother(2): . . . TCAAATCCTTTTAATAAAGAAGAGCTGACA . . .

Sibling(1): . . . TCAAATCCTTTTAATAAAGAAGAGCTGACA . . .

Sibling(2): . . . TCAAATCCTTTTAATAAAGAAGAGCTGACA . . .

Proband(1): . . . TCAAATCCTTTTAATAAAGAAGAGCTGACA . . .

Proband(2): . . . TCAAATCCTTTTAAT\*\*\*\*AAGAGCTGACA . . .

4bp heterozygous deletion at chr15:9352406 | CHD2

# De novo Genetics of Autism

- In 593 family quads so far, we see significant enrichment in de novo **likely gene disruptions (LGDs)** in the autistic kids
  - Overall rate basically 1:1
  - 2:1 enrichment in frameshift indels (35:16)
- Confirmed trends observed in previous studies, contributed dozens of new autism candidate genes.
  - 8 out of 35 indel LGDs in autistic children overlapped with the 842 FMRP-associated genes
  - Trends further confirmed in larger study over the entire collection that is currently under review

**Accurate de novo and transmitted indel detection in exome-capture data using microassembly.**  
Narzisi et al. (2014) *Nature Methods* doi:10.1038/nmeth.3069

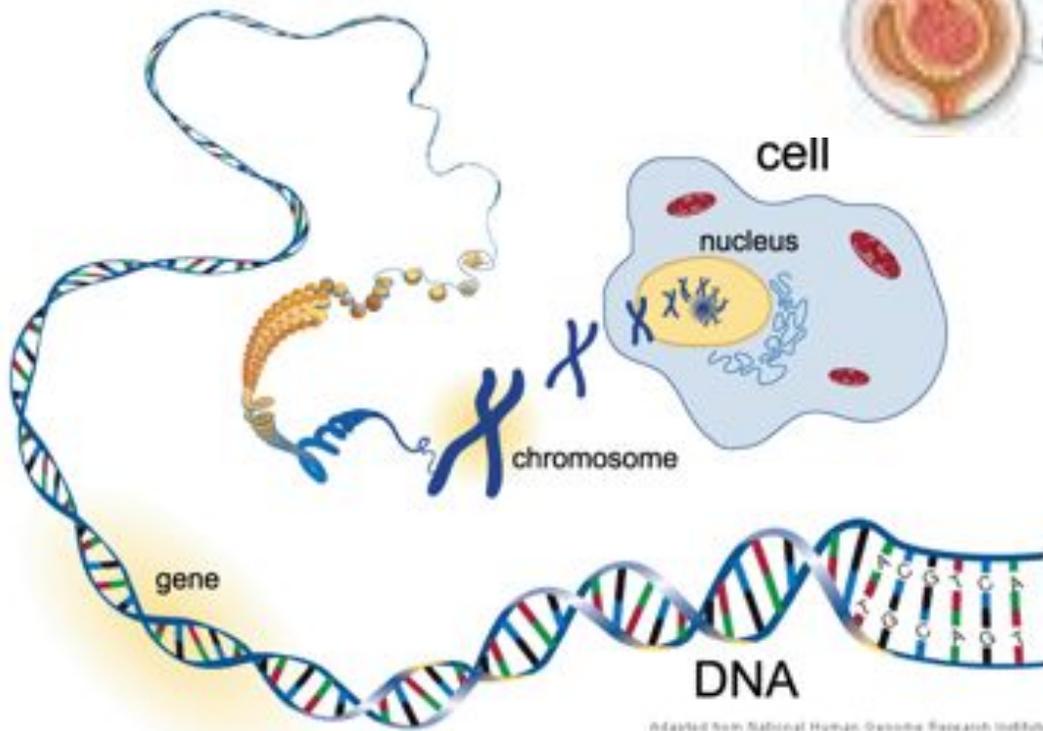
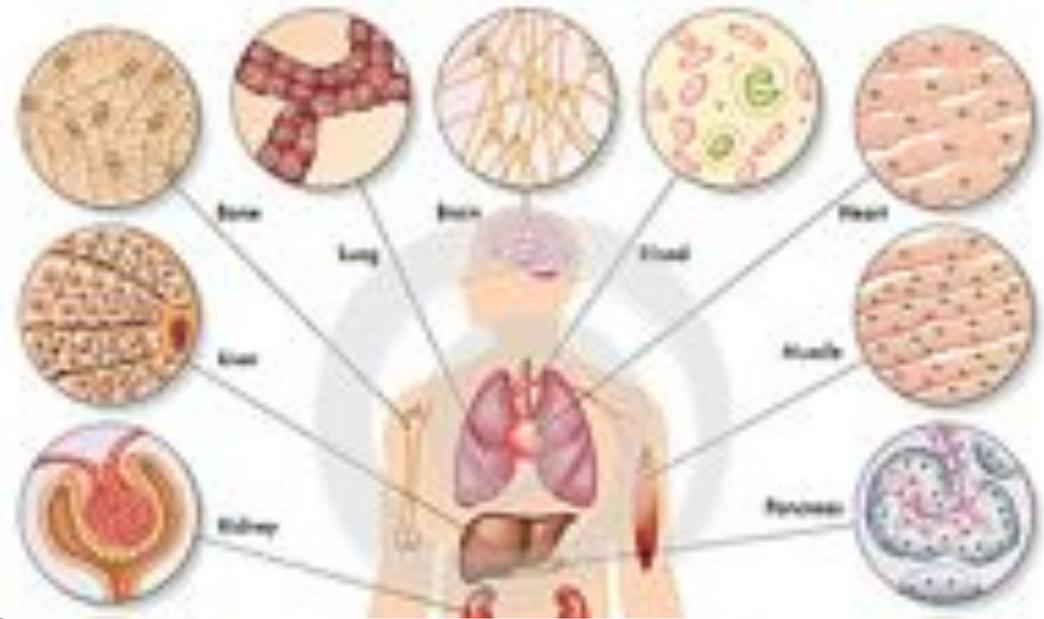
**The burden of de novo coding mutations in autism spectrum disorders.**  
Iossifov et al (2014) *Under review.*



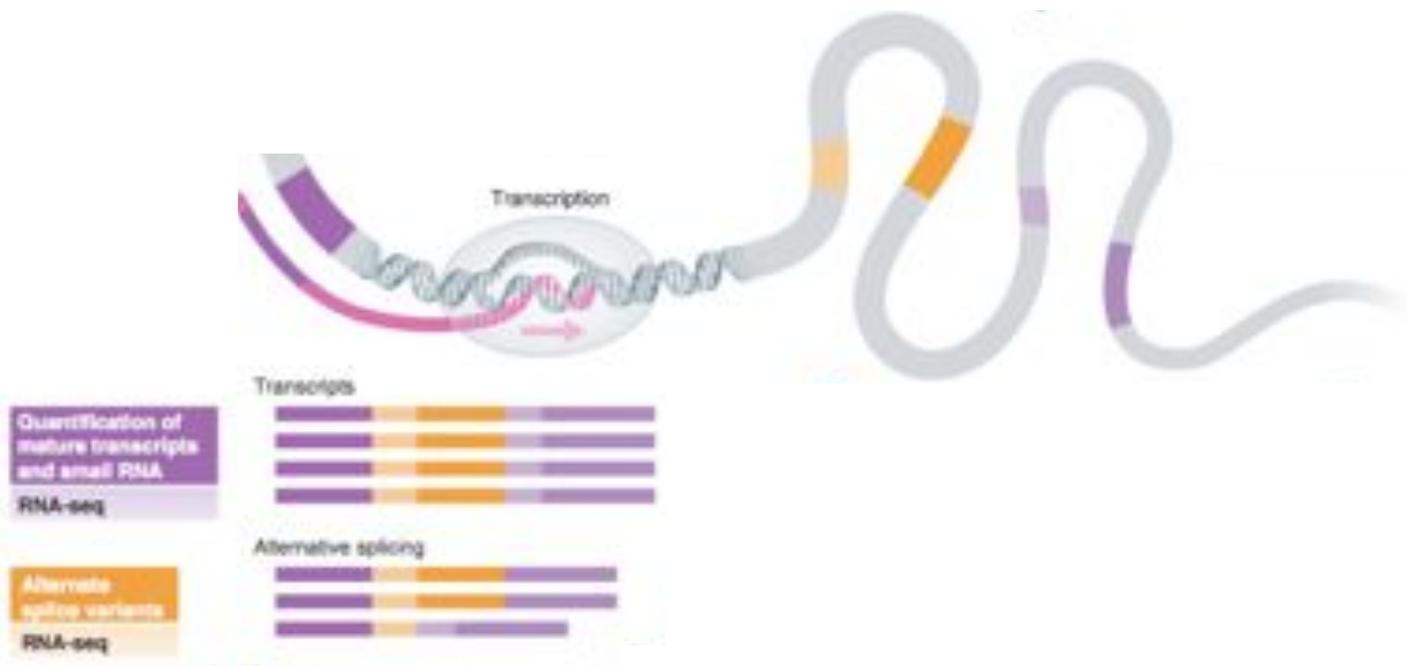
Break

# Cells & DNA

Each cell of your body contains an exact copy of your 3 billion base pair genome.



Your specific nucleotide sequence encodes the genetic program for your cells and ultimately your traits



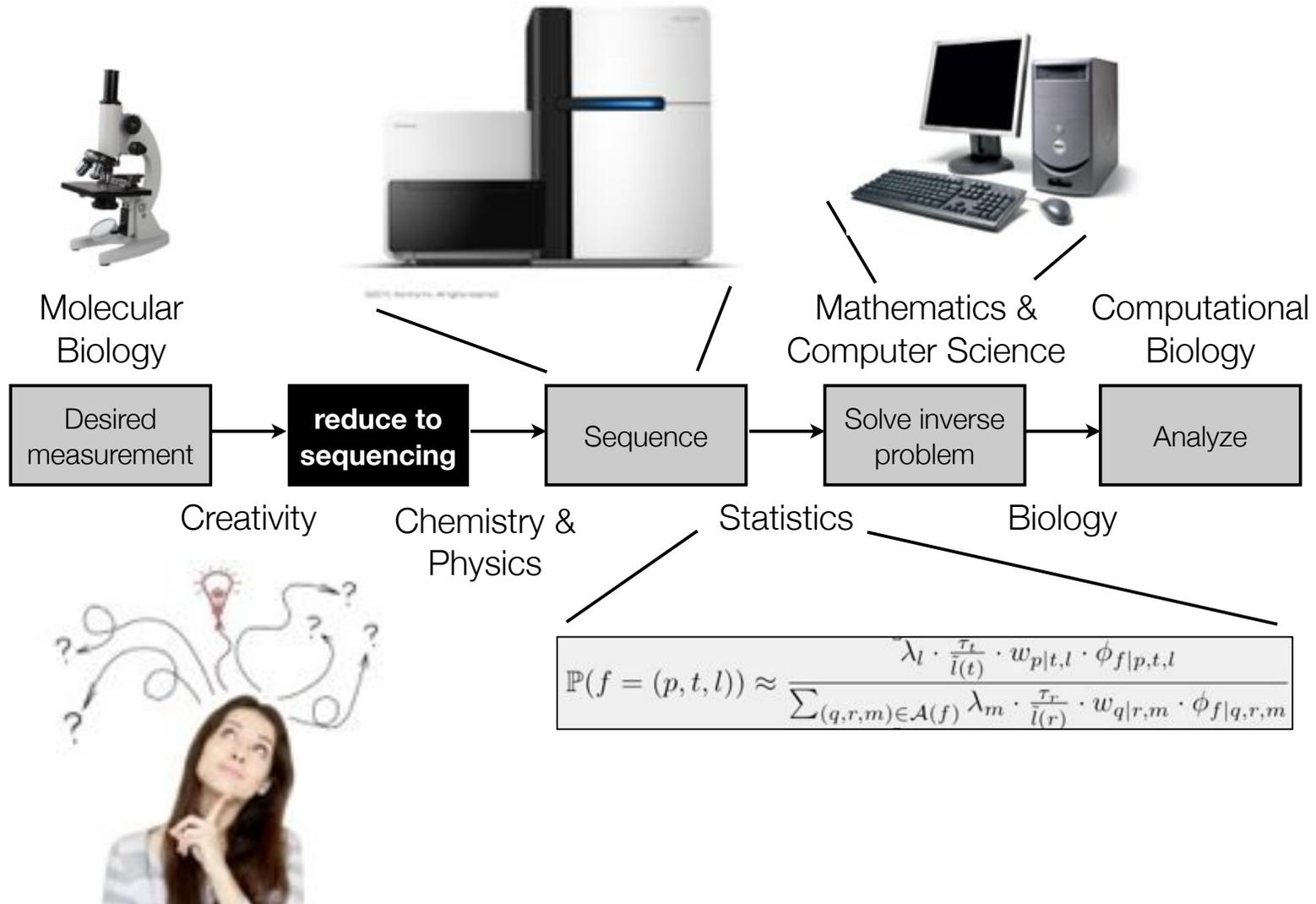
Soon et al., Molecular Systems Biology, 2013

# Sequencing Assays

## The \*Seq List (in chronological order)

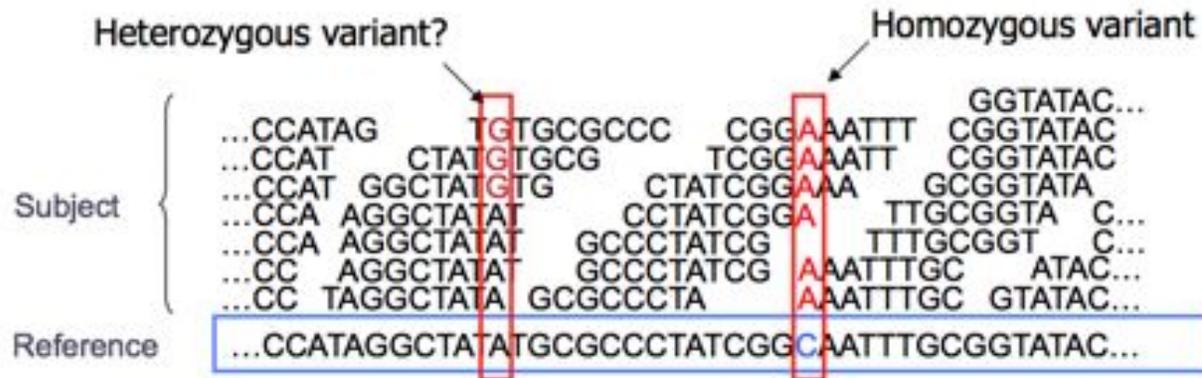
1. Gregory E. Crawford et al., "Genome-wide Mapping of DNase Hypersensitive Sites Using Massively Parallel Signature Sequencing (MPSS)," *Genome Research* 16, no. 1 (January 1, 2006): 123–131, doi:10.1101/gr.4074106.
2. David S. Johnson et al., "Genome-Wide Mapping of in Vivo Protein-DNA Interactions," *Science* 316, no. 5830 (June 8, 2007): 1497–1502, doi:10.1126/science.1141319.
3. Tarjei S. Mikkelsen et al., "Genome-wide Maps of Chromatin State in Pluripotent and Lineage-committed Cells," *Nature* 448, no. 7153 (August 2, 2007): 553–560, doi:10.1038/nature06008.
4. Thomas A. Down et al., "A Bayesian Deconvolution Strategy for Immunoprecipitation-based DNA Methylome Analysis," *Nature Biotechnology* 26, no. 7 (July 2008): 779–785, doi:10.1038/nbt1414.
5. Ali Mortazavi et al., "Mapping and Quantifying Mammalian Transcriptomes by RNA-Seq," *Nature Methods* 5, no. 7 (July 2008): 621–628, doi:10.1038/nmeth.1226.
6. Nathan A. Baird et al., "Rapid SNP Discovery and Genetic Mapping Using Sequenced RAD Markers," *PLoS ONE* 3, no. 10 (October 13, 2008): e3376, doi:10.1371/journal.pone.0003376.
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11. Alayne L. Brunner et al., "Distinct DNA Methylation Patterns Characterize Differentiated Human Embryonic Stem Cells and Developing Human Fetal Liver," *Genome Research* 19, no. 6 (June 1, 2009): 1044–1056, doi:10.1101/gr.088773.108.
12. Mayumi Oda et al., "High-resolution Genome-wide Cytosine Methylation Profiling with Simultaneous Copy Number Analysis and Optimization for Limited Cell Numbers," *Nucleic Acids Research* 37, no. 12 (July 1, 2009): 3829–3839, doi:10.1093/nar/gkp260.
13. Zachary D. Smith et al., "High-throughput Bisulfite Sequencing in Mammalian Genomes," *Methods* 48, no. 3 (July 2009): 226–232, doi:10.1016/j.ymeth.2009.05.003.
14. Andrew M. Smith et al., "Quantitative Phenotyping via Deep Barcode Sequencing," *Genome Research* (July 21, 2009), doi:10.1101/gr.

# What is a \*Seq assay?

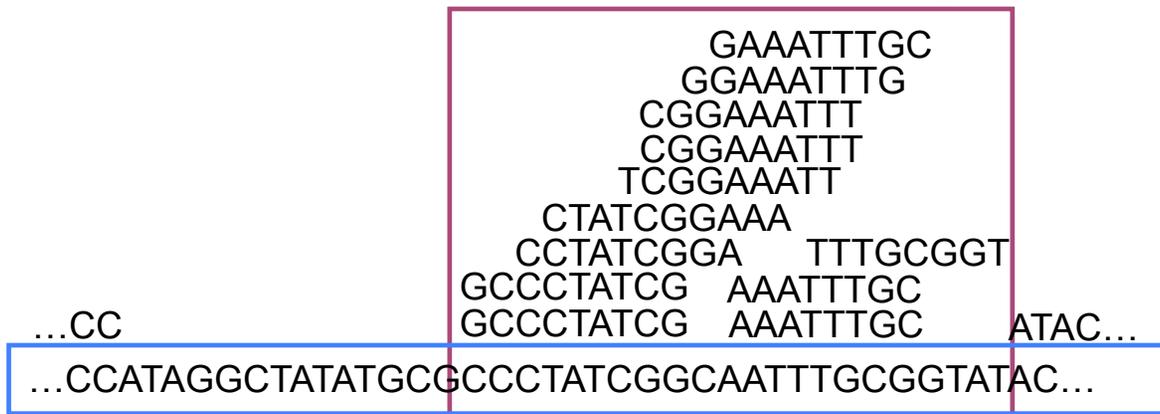


# Short Read Applications

- Genotyping: Identify Variations

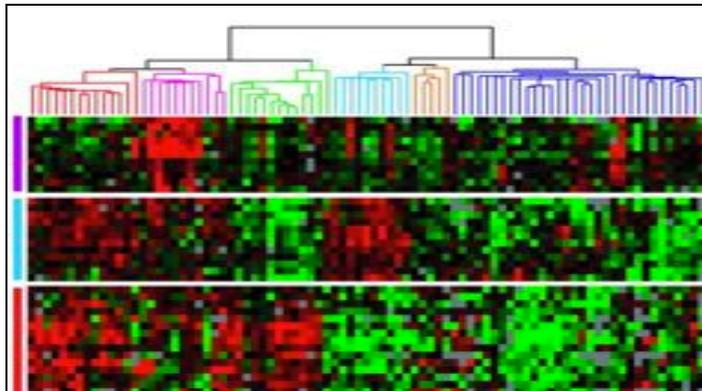


- \*-seq: Classify & measure significant peaks

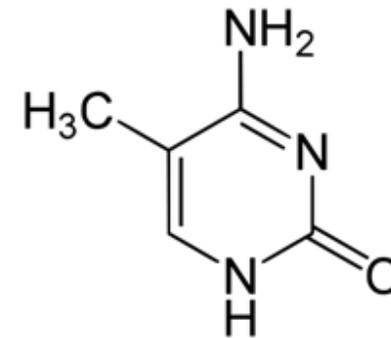


# \*-seq in 4 short vignettes

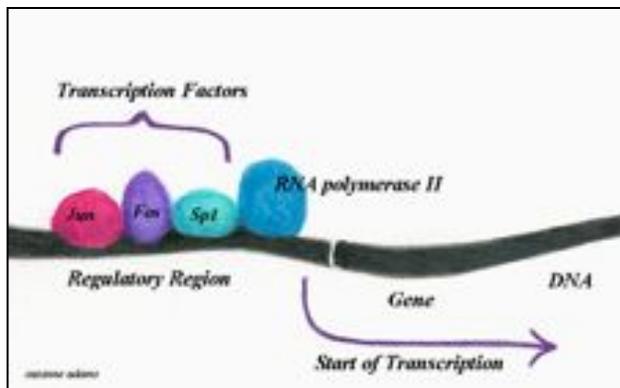
## RNA-seq



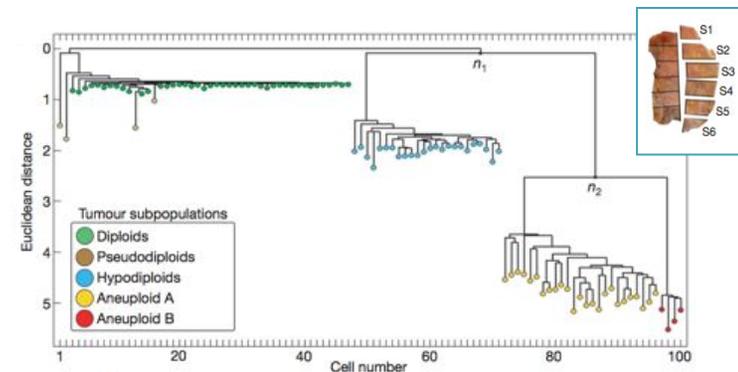
## Methyl-seq



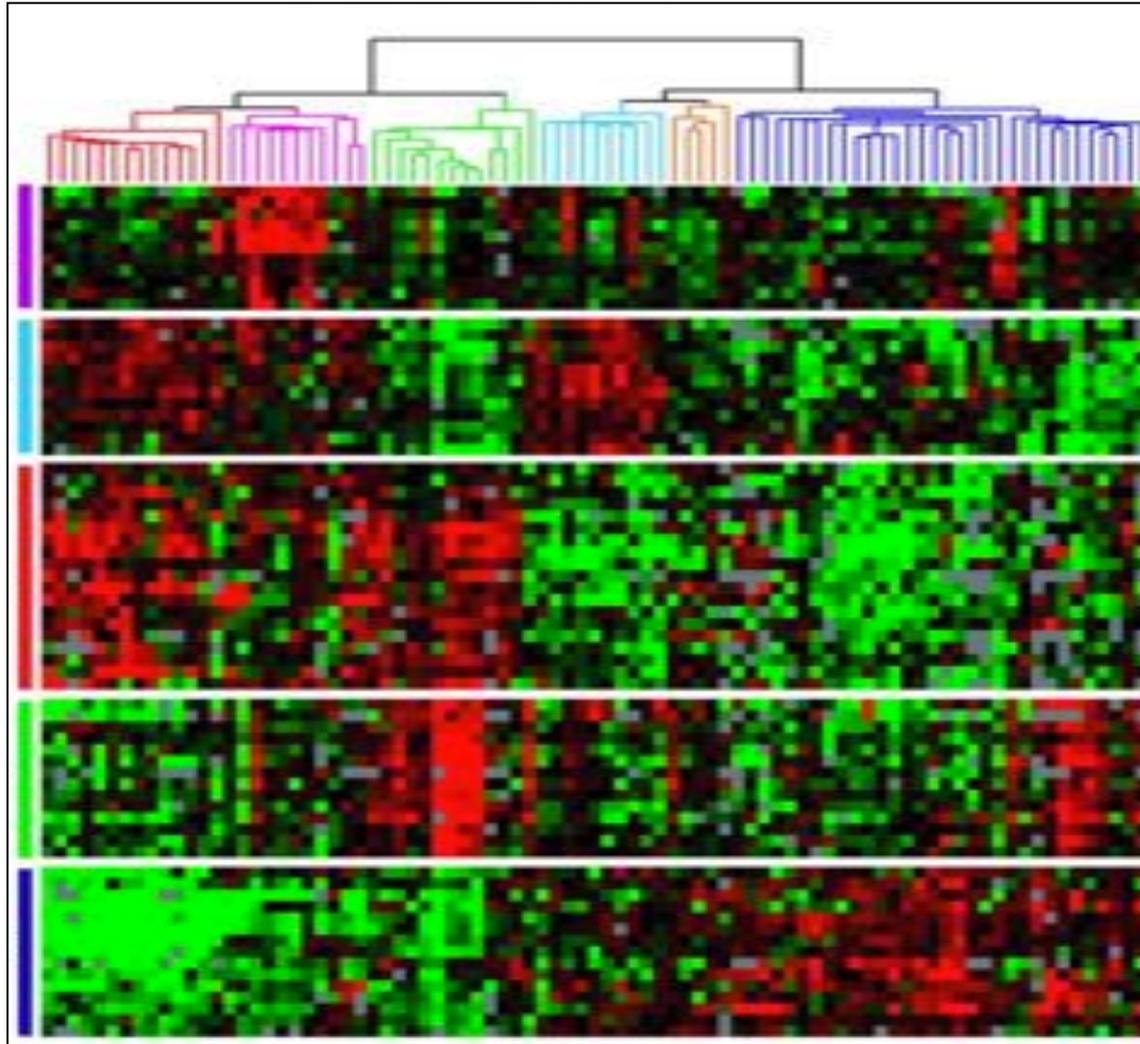
## ChIP-seq



## Single Cell-seq

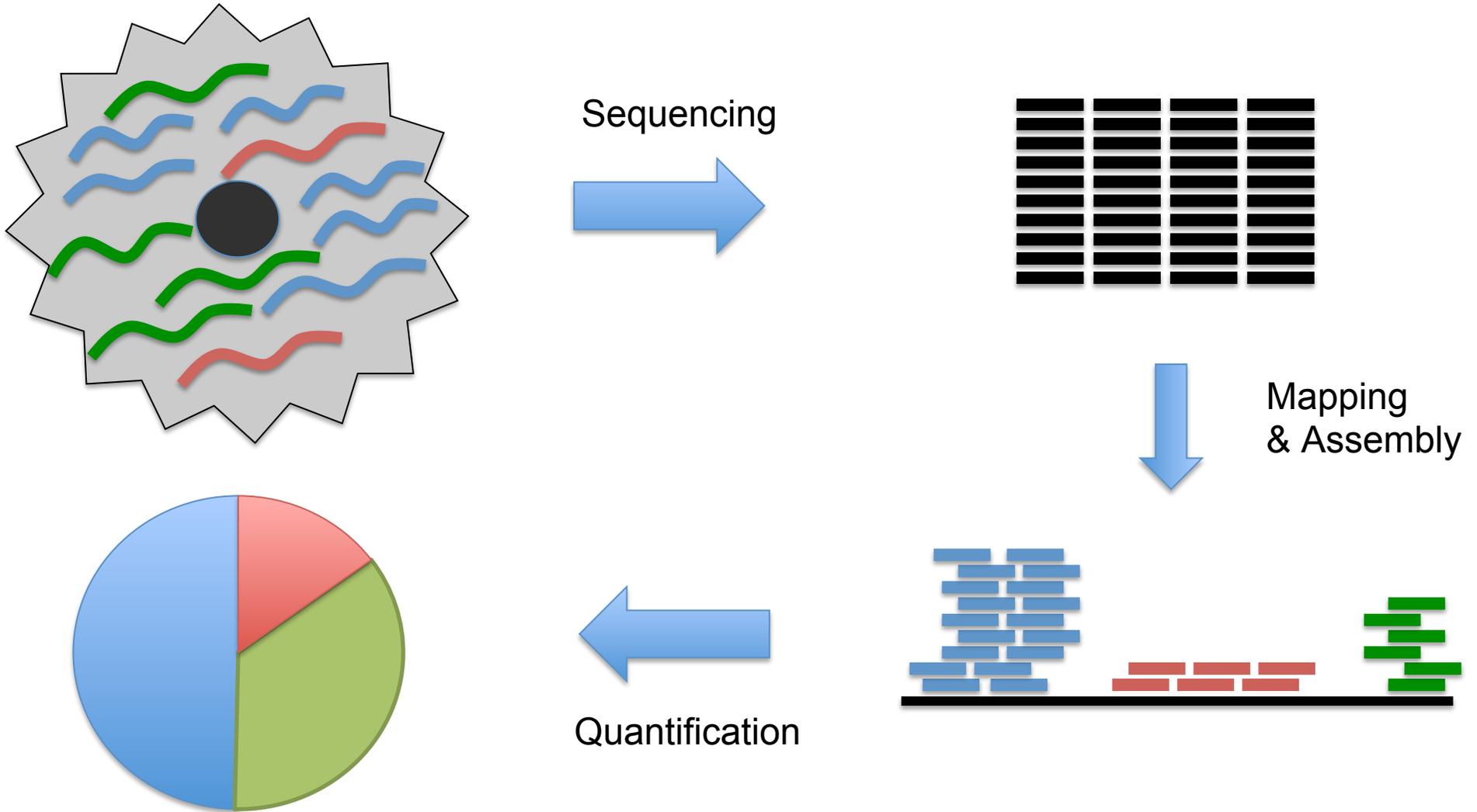


# RNA-seq

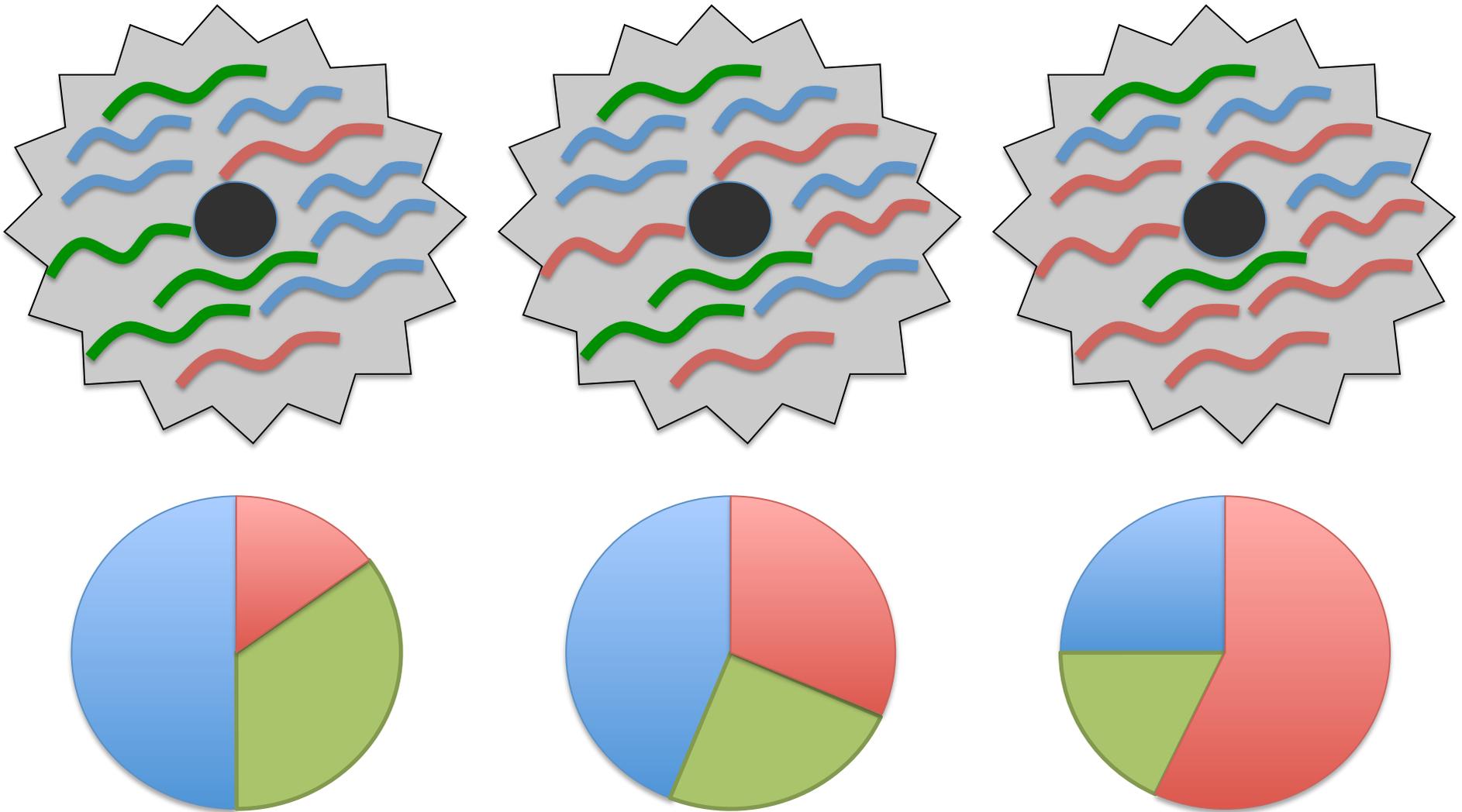


**Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications.**  
Sørlie et al (2001) *PNAS*. 98(19):10869-74.

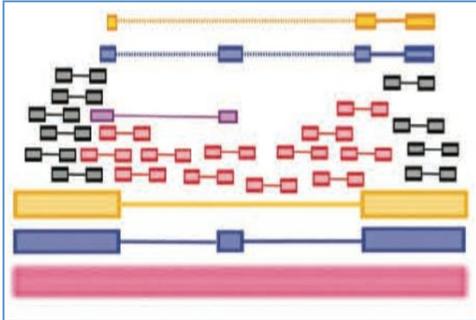
# RNA-seq Overview



# RNA-seq Overview



# RNA-seq Challenges

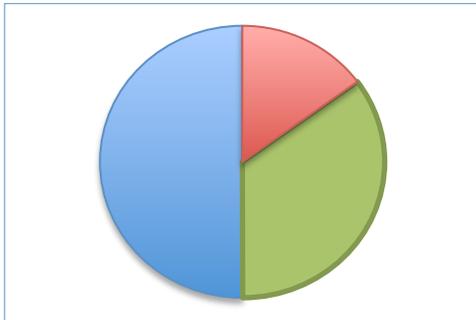


## Challenge 1: Eukaryotic genes are spliced

Solution: Use a spliced aligner, and assemble isoforms

### TopHat: discovering spliced junctions with RNA-Seq.

Trapnell et al (2009) *Bioinformatics*. 25:0 | 105-111

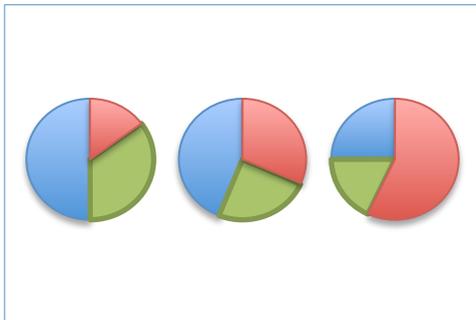


## Challenge 2: Read Count $\neq$ Transcript abundance

Solution: Infer underlying abundances (e.g. FPKM)

### Transcript assembly and quantification by RNA-seq

Trapnell et al (2010) *Nat. Biotech.* 25(5): 511-515



## Challenge 3: Transcript abundances are stochastic

Solution: Replicates, replicates, and more replicates

### RNA-seq differential expression studies: more sequence or more replication?

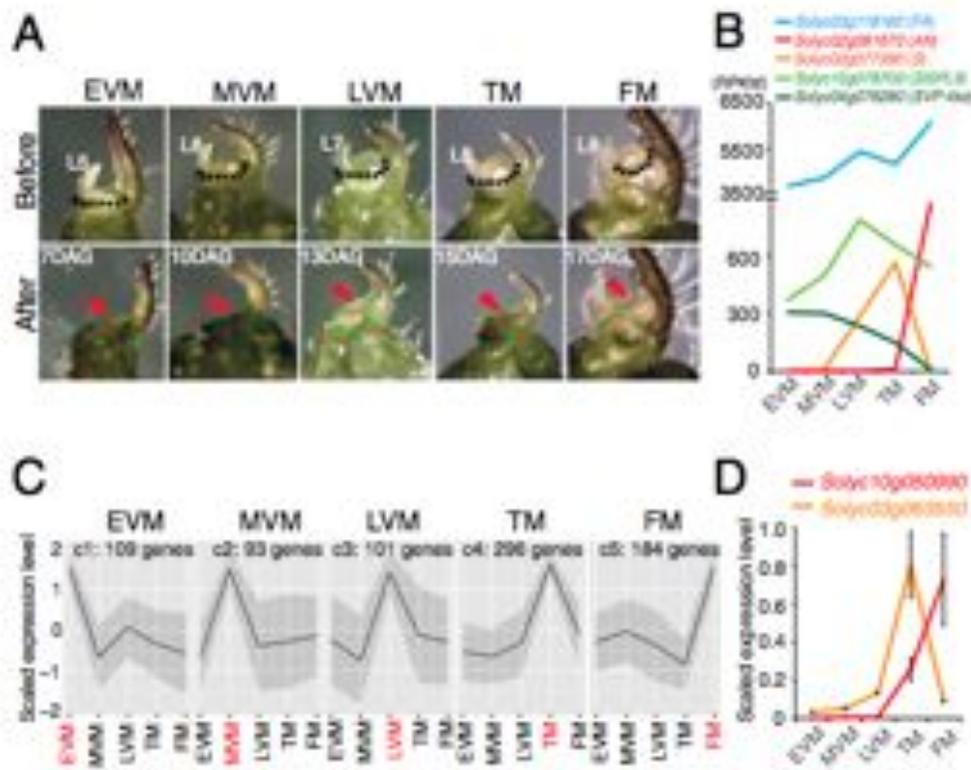
Liu et al (2013) *Bioinformatics*. doi:10.1093/bioinformatics/btt688

# Rate of meristem maturation determines inflorescence architecture in tomato

Soon Ju Park<sup>1</sup>, Ke Jiang<sup>1</sup>, Michael C. Schatz, and Zachary B. Lippman<sup>2</sup>

Cold Spring Harbor Laboratory, Cold Spring Harbor, NY 11724

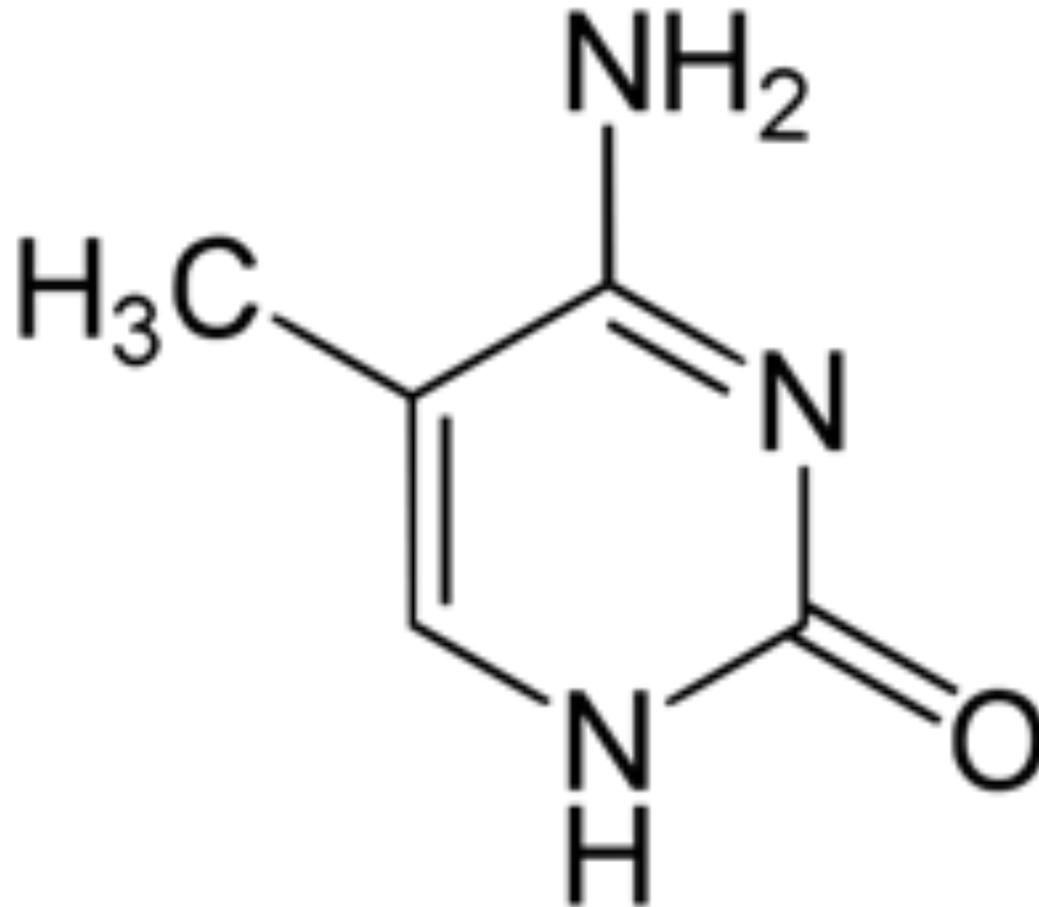
Edited by Maarten Koornneef, Wageningen University and Research Centre, Cologne, Germany, and approved November 28, 2011 (received for review September 12, 2011)



## RNA-seq to determine the expression dynamics during development

- Laser microdissection to precisely extract tissue from developing organs
- Use RNA-seq to watch different classes of genes become activated at different stages of development
- When those genes are delayed or interrupted, tomato mutants take on very different branching patterns.

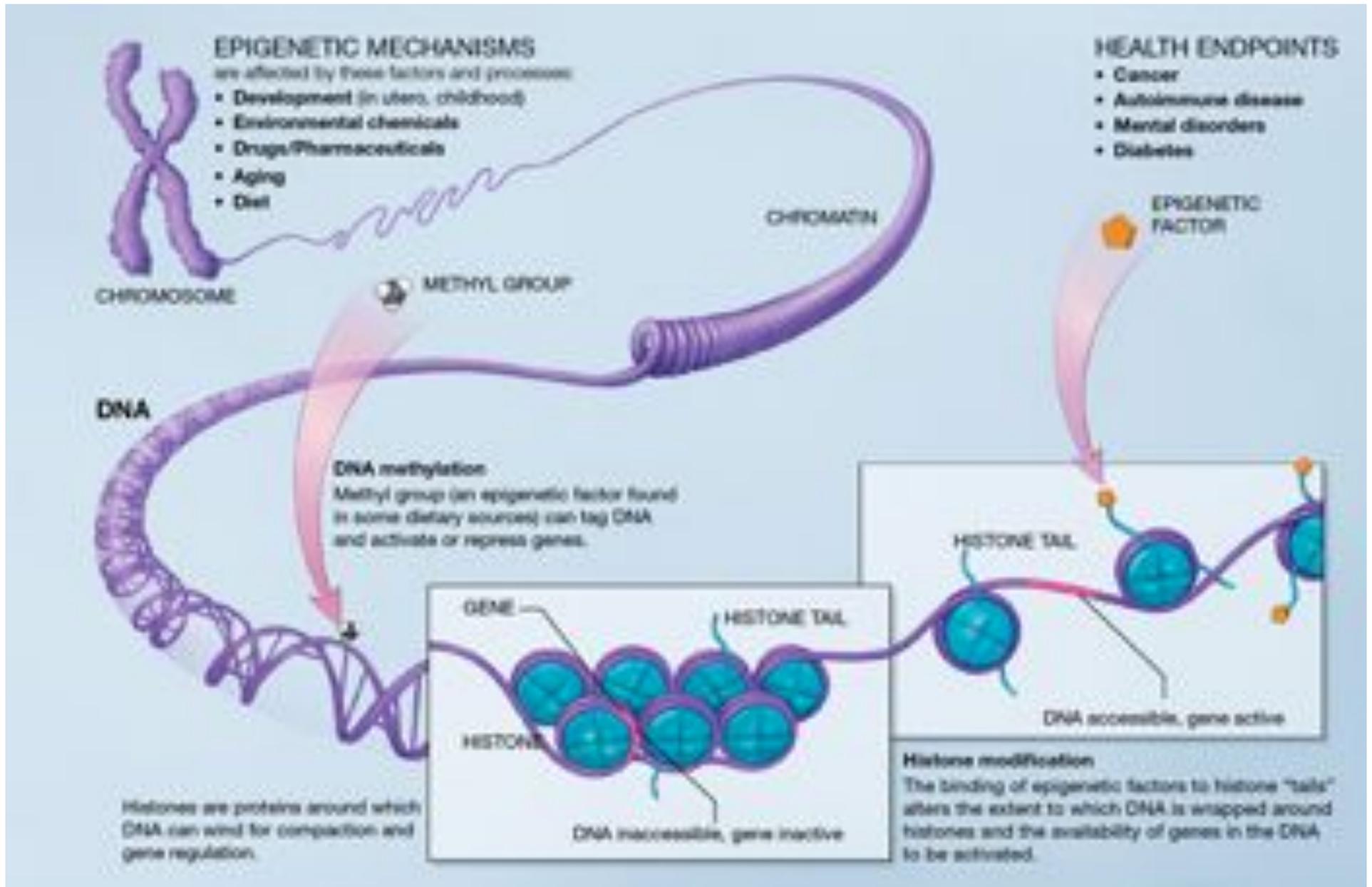
# Methyl-seq



**Finding the fifth base: Genome-wide sequencing of cytosine methylation**

Lister and Ecker (2009) *Genome Research*. 19: 959-966

# Methylation & Epigenetics



# The Honey Bee Epigenomes: Differential Methylation of Brain DNA in Queens and Workers

Frank Lyko<sup>1\*</sup>, Sylvain Foret<sup>2,3</sup>, Robert Kucharski<sup>3</sup>, Stephan Wolf<sup>4</sup>, Cassandra Falckenhayn<sup>1</sup>, Ryszard Maleszka<sup>3\*</sup>

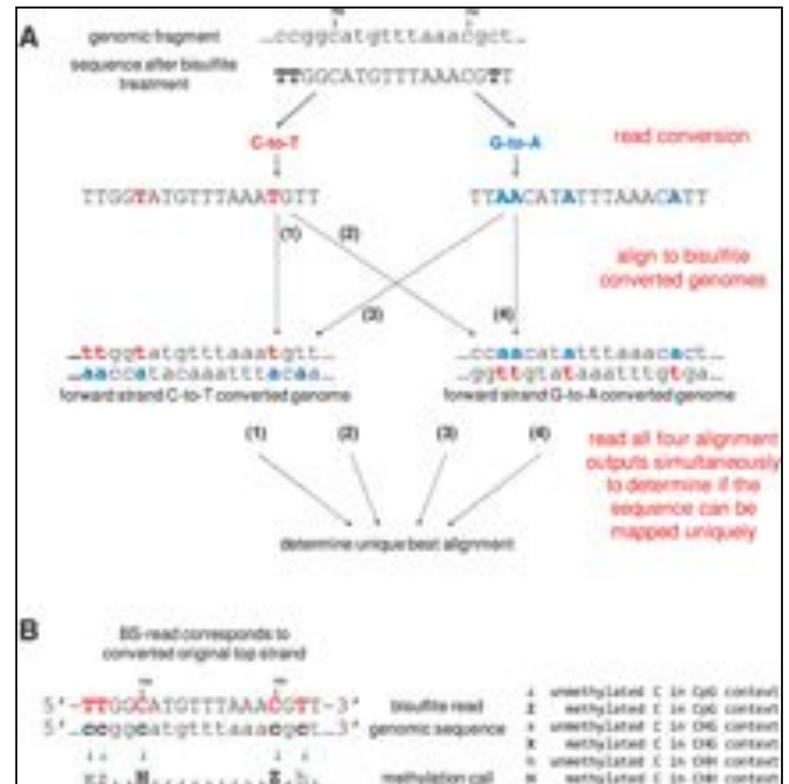
**1** Division of Epigenetics, DKFZ-ZMBH Alliance, German Cancer Research Center, Heidelberg, Germany, **2** ARC Centre of Excellence for Coral Reef Studies, James Cook University, Townsville, Australia, **3** Research School of Biology, the Australian National University, Canberra, Australia, **4** Genomics and Proteomics Core Facility, German Cancer Research Center, Heidelberg, Germany



# Bisulfite Conversion

## Treating DNA with sodium bisulfite will convert un methylated C to T

- 5-MethyC will be protected and not change, so can look for differences when mapping
- Requires great care when analyzing reads, since the complementary strand will also be converted (G to A)
- Typically analyzed by mapping to a “reduced alphabet” where we assume all Cs are converted to Ts once on the forward strand and once on the reverse



## Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications

Krueger and Andrews (2010) *Bioinformatics*. 27 (11): 1571-1572.

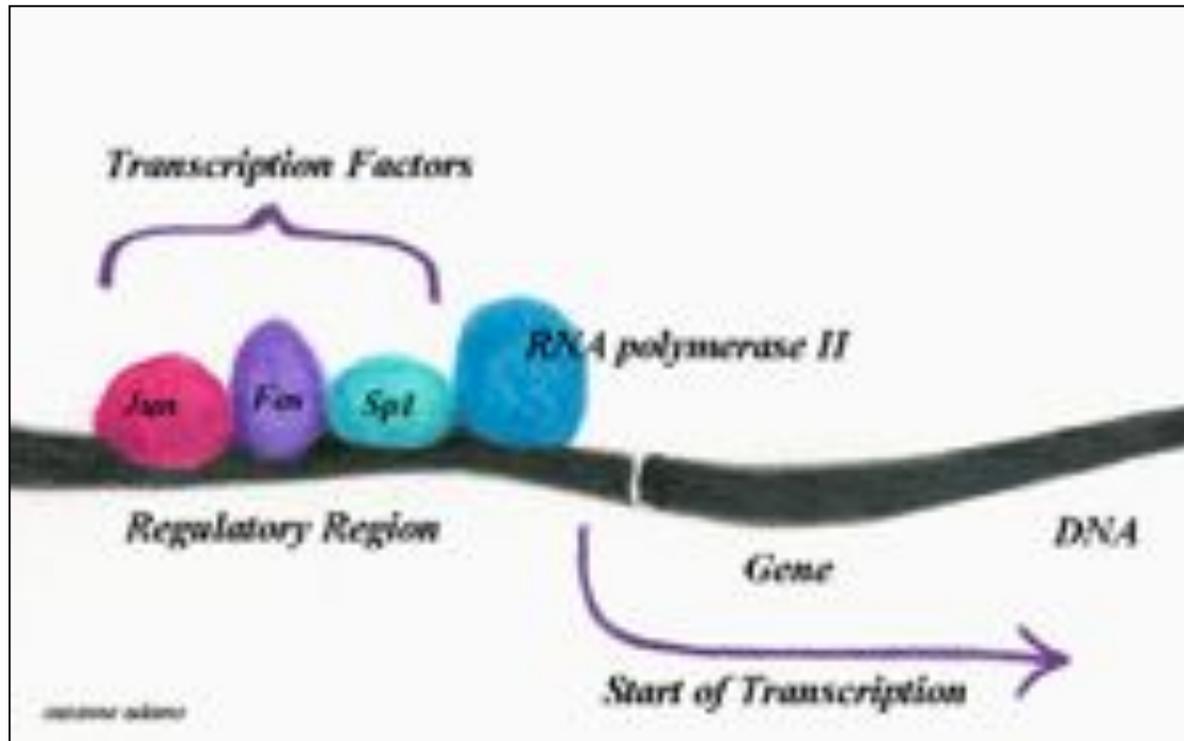
# Bisulfite Conversion

T  
W  
•  
•  
•



**Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications**  
Krueger and Andrews (2010) *Bioinformatics*. 27 (11): 1571-1572.

# ChIP-seq



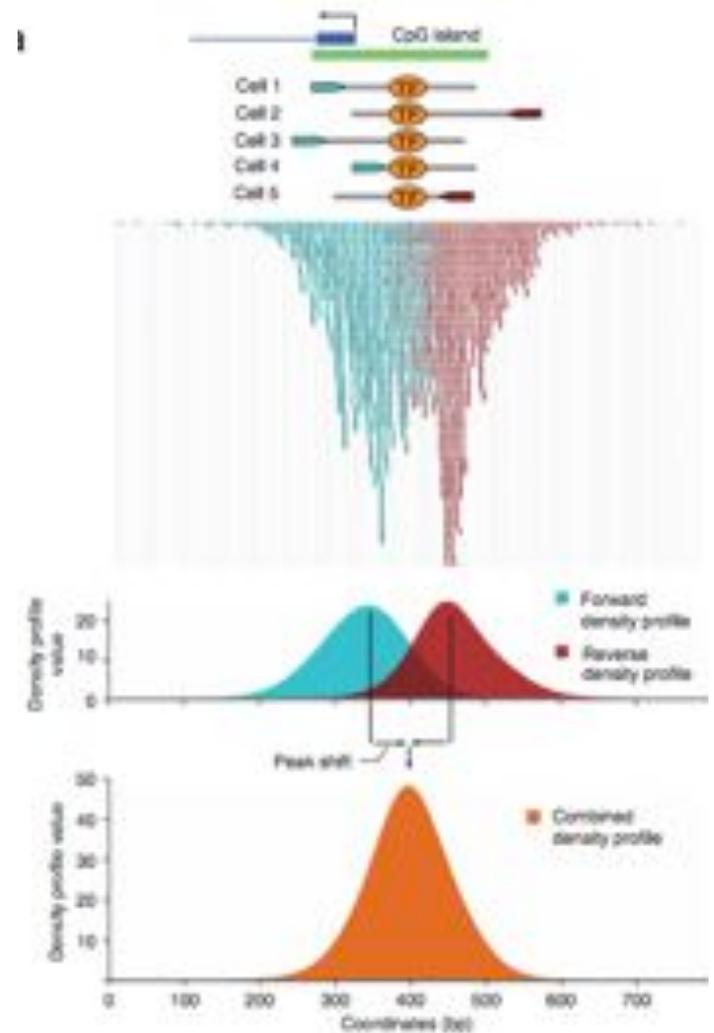
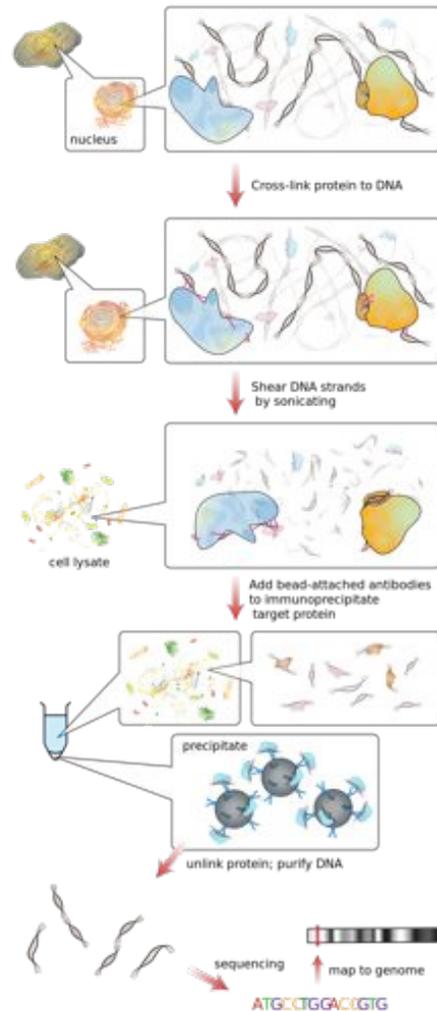
**Genome-wide mapping of in vivo protein-DNA interactions.**

Johnson *et al* (2007) *Science*. 316(5830):1497-502

# ChIP-seq

## Goals:

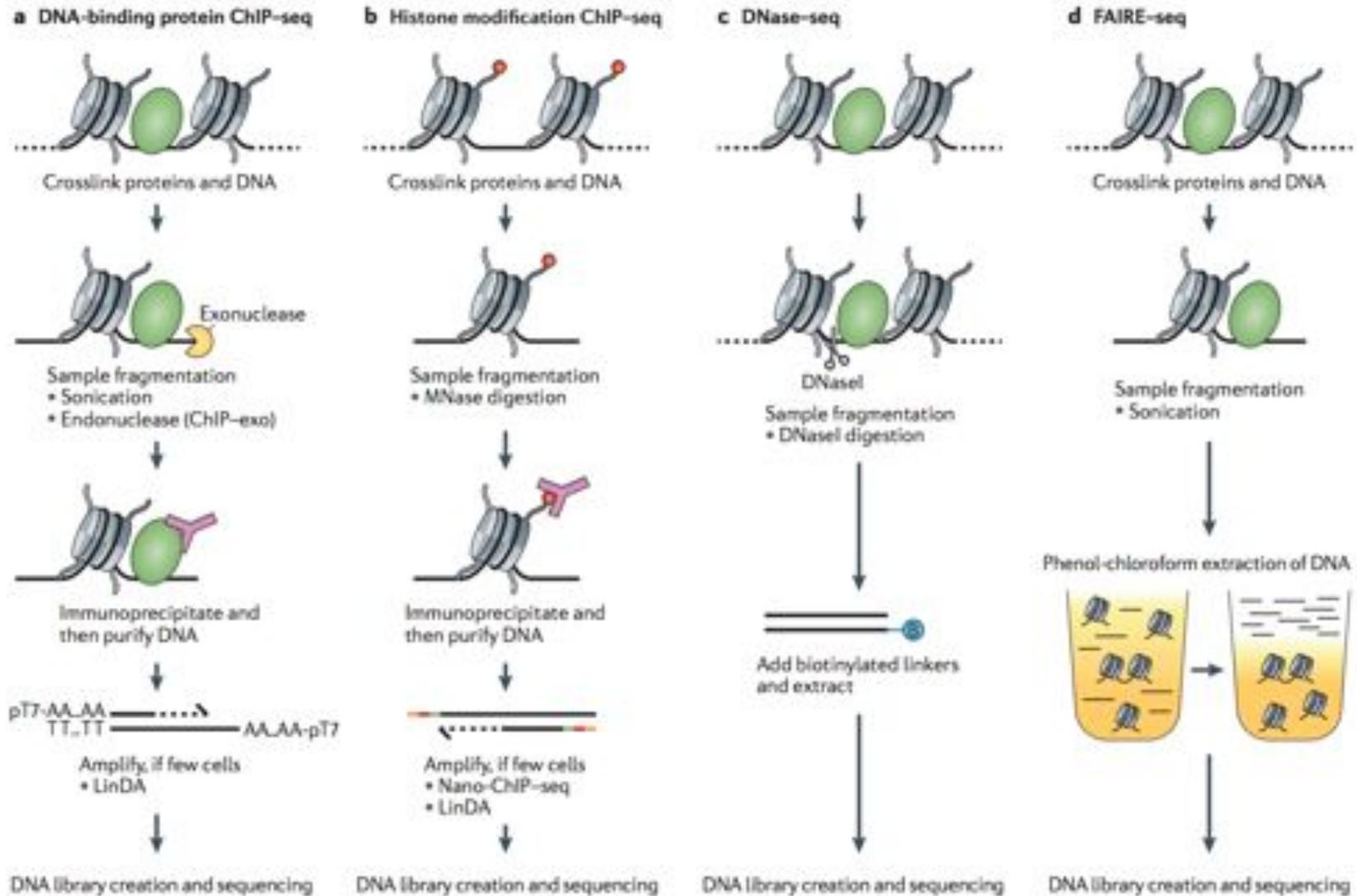
- Where are transcription factors and other proteins binding to the DNA?
- How strongly are they binding?
- Do the protein binding patterns change over developmental stages or when the cells are stressed?



## Genome-wide analysis of transcription factor binding sites based on ChIP-Seq data

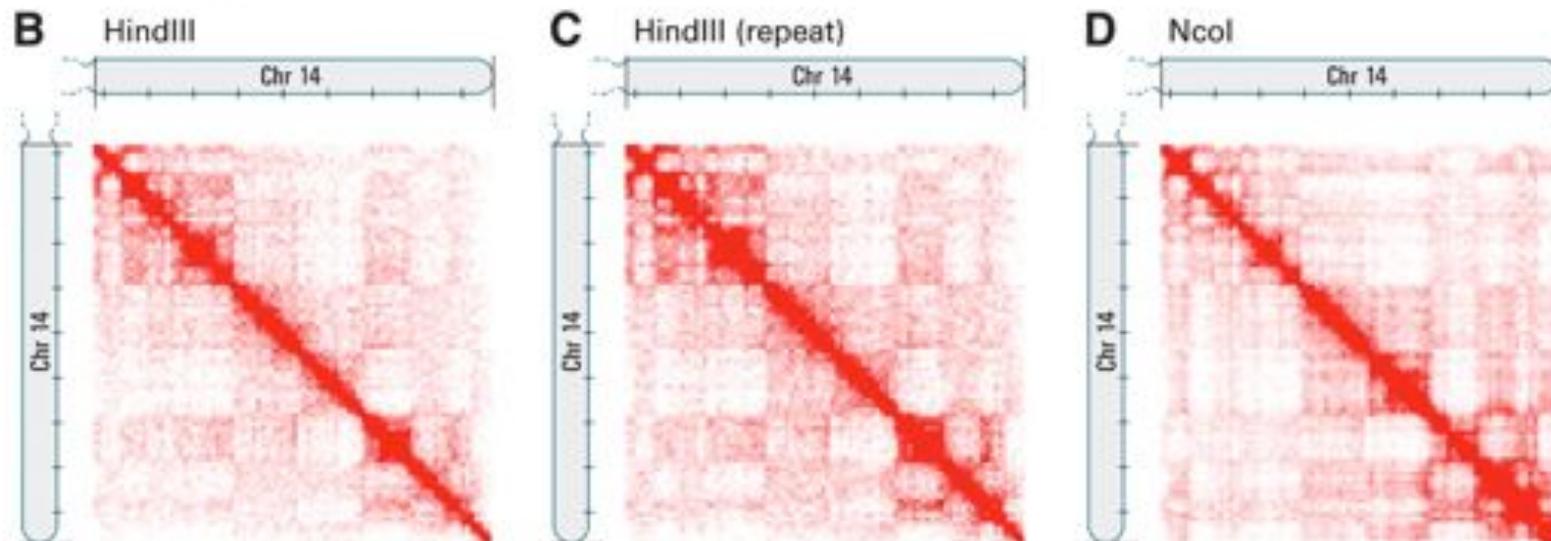
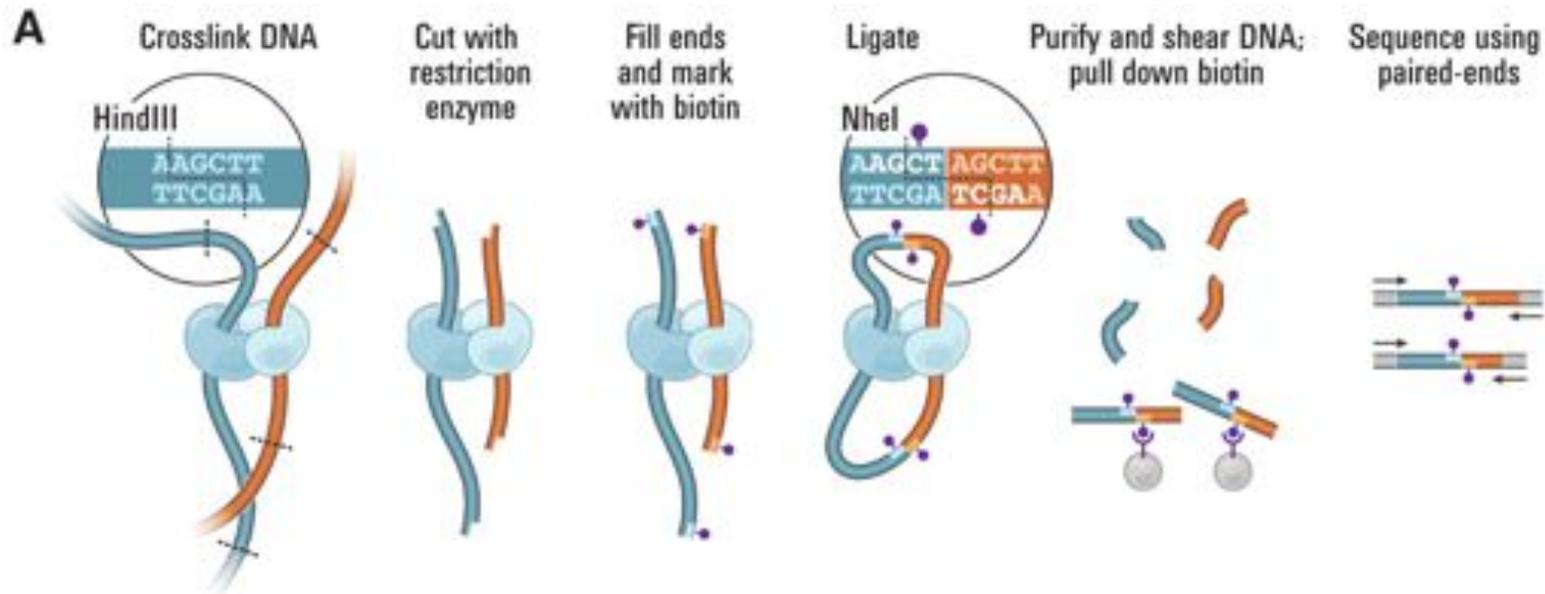
Valouev et al (2008) *Nature Methods*. 5, 829 - 834

# Related Assays



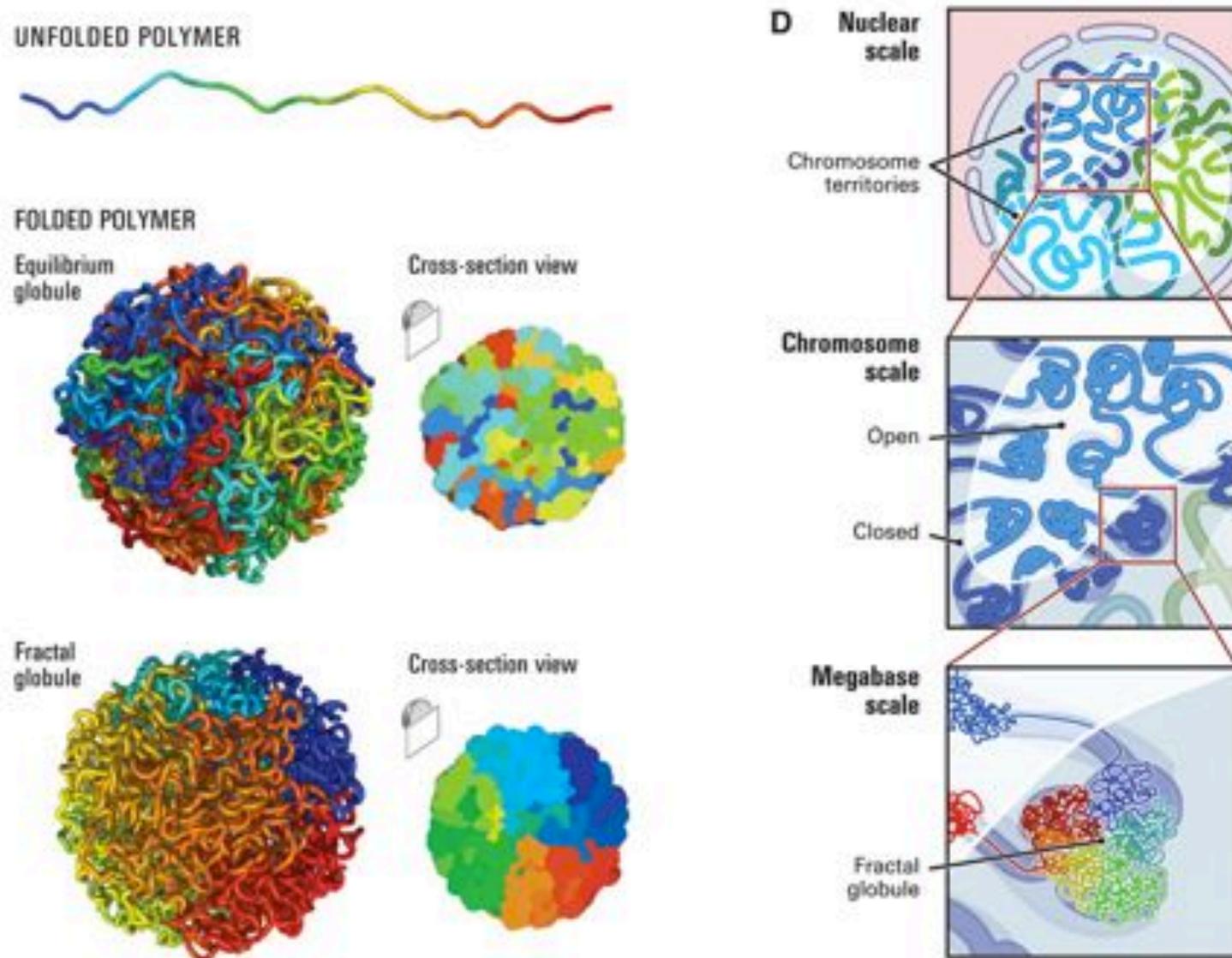
**ChIP-seq and beyond: new and improved methodologies to detect and characterize protein-DNA interactions**  
 Furey (2012) *Nature Reviews Genetics*. 13, 840-852

# Hi-C: Mapping the folding of DNA



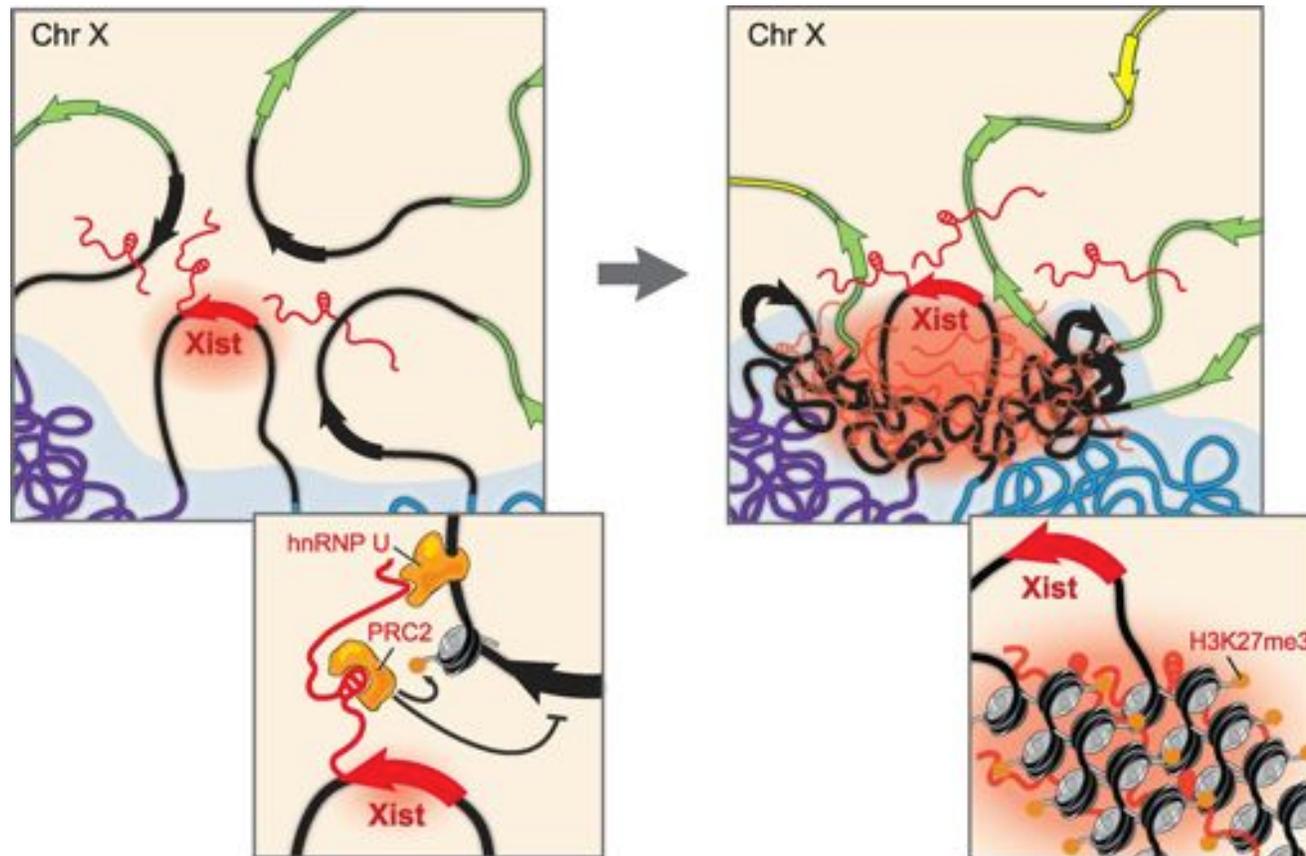
**Comprehensive Mapping of Long-Range Interactions Reveals Folding Principles of the Human Genome**  
Lieberman-Aiden et al. (2009) *Science*. 326 (5950): 289-293

# Hi-C: Mapping the folding of DNA



**Comprehensive Mapping of Long-Range Interactions Reveals Folding Principles of the Human Genome**  
Lieberman-Aiden et al. (2009) *Science*. 326 (5950): 289-293

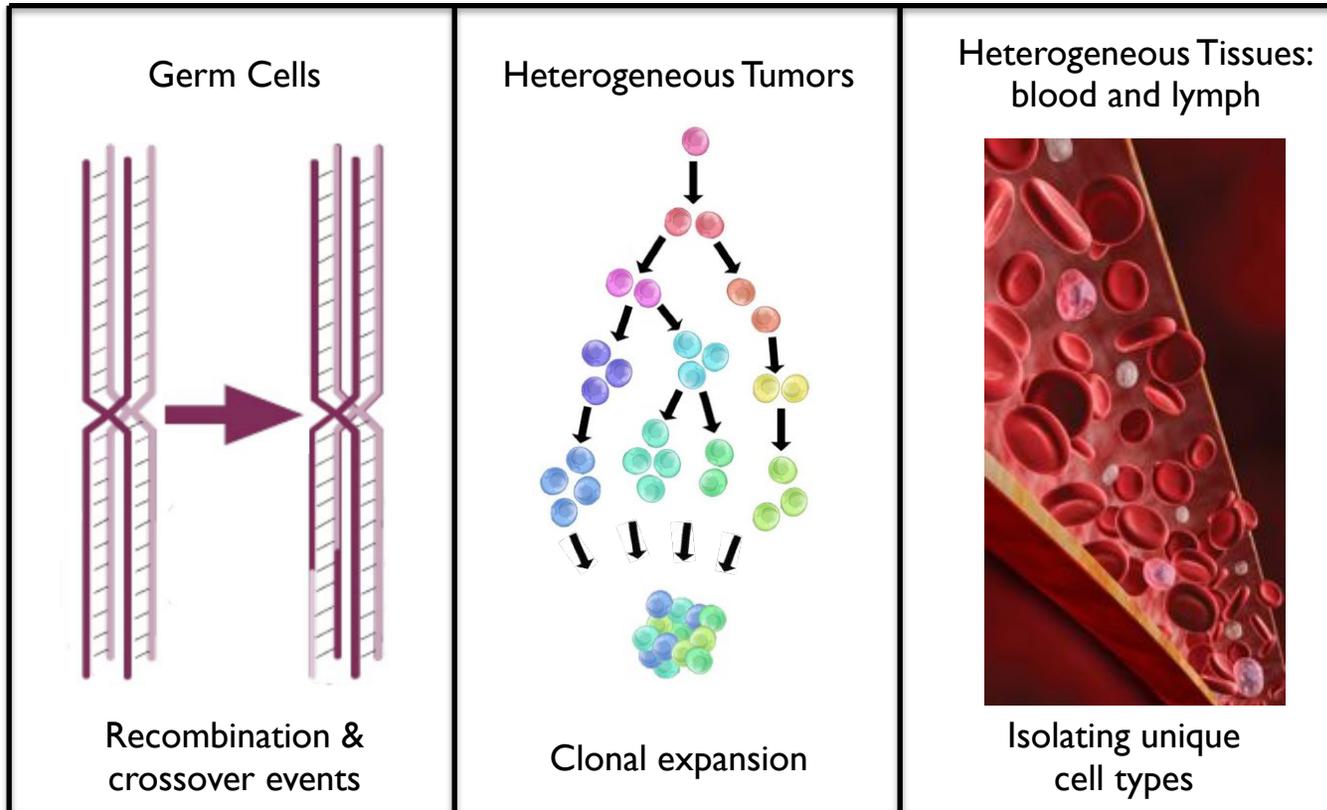
# Gene Regulation in 3-dimensions



**Fig 6. A model for how Xist exploits and alters three-dimensional genome architecture to spread across the X chromosome.**

The Xist lncRNA Exploits Three-Dimensional Genome Architecture to Spread Across the X Chromosome  
Engreitz et al. (2013) *Science*. 341 (6147)

# Single Cell Sequencing



**Cancer genomics: one cell at a time**

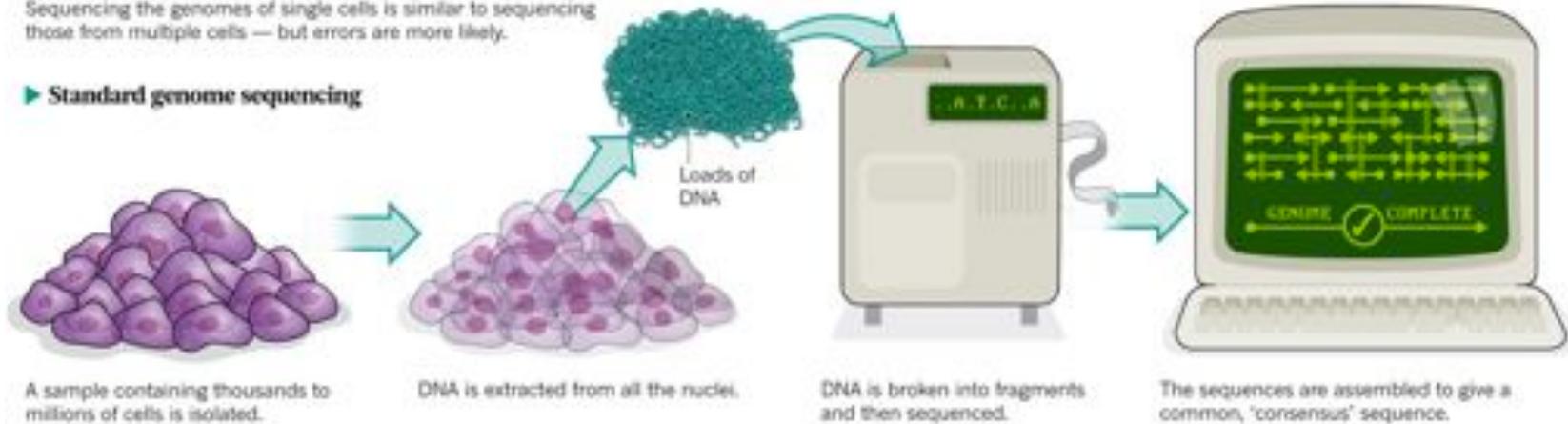
Navin et al (2014) *Genome Biology*. 15:452

# Bulk vs Single Cell

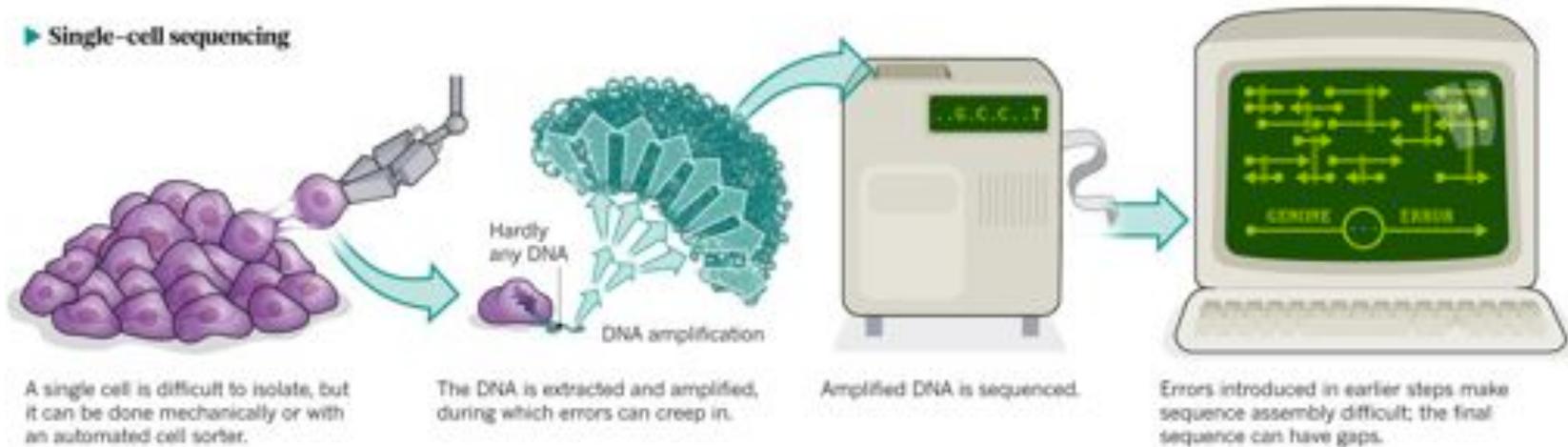
## ONE GENOME FROM MANY

Sequencing the genomes of single cells is similar to sequencing those from multiple cells — but errors are more likely.

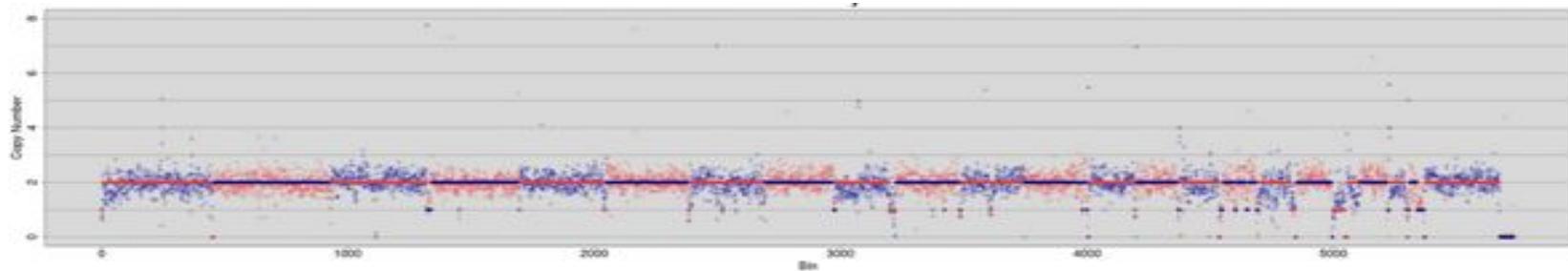
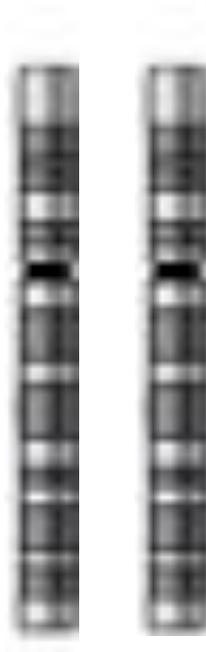
### ► Standard genome sequencing



### ► Single-cell sequencing

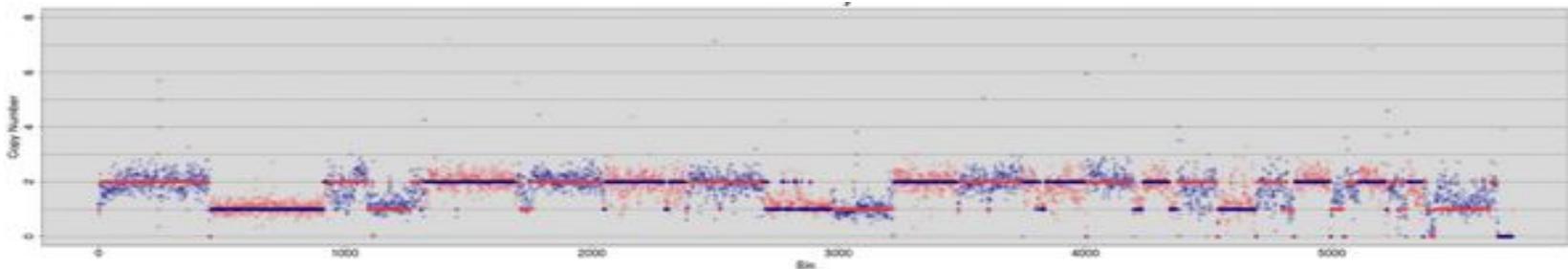
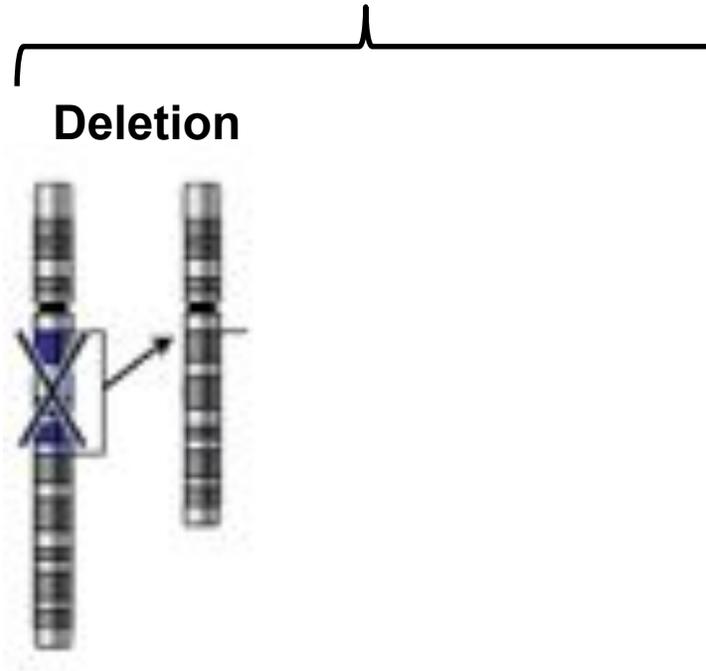


# Copy Number Variants



# Copy Number Variants

## Structural Variation

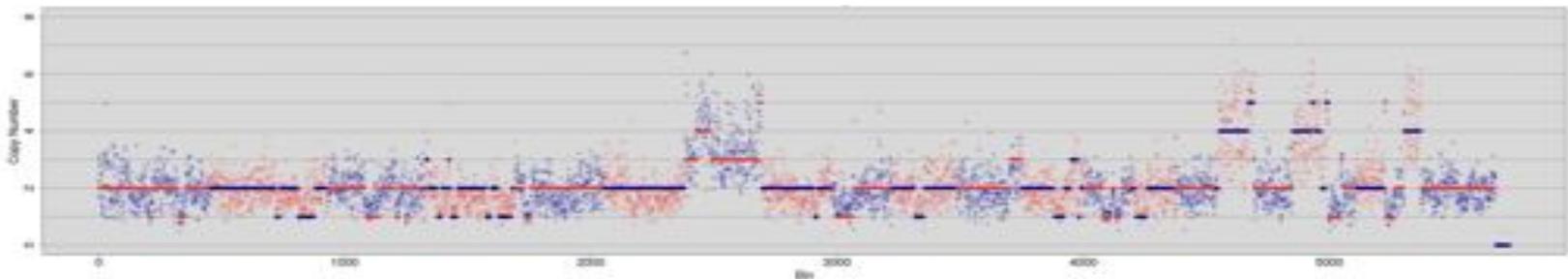
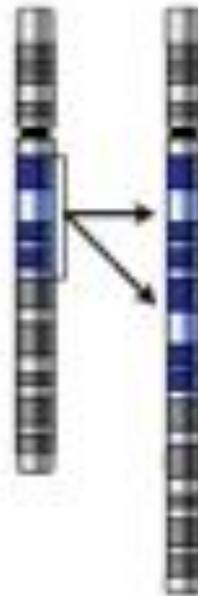


# Copy Number Variants

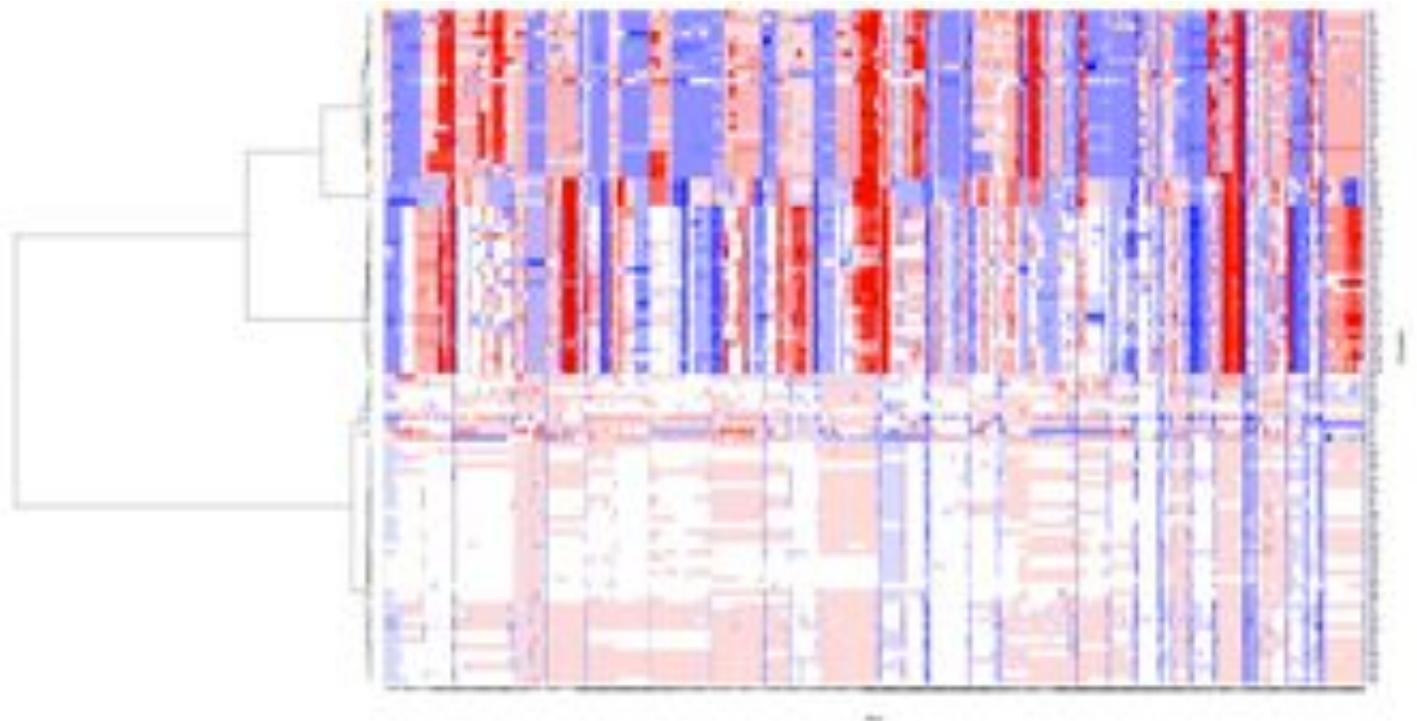
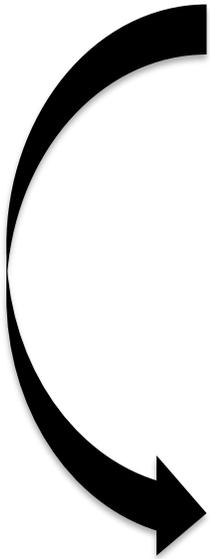
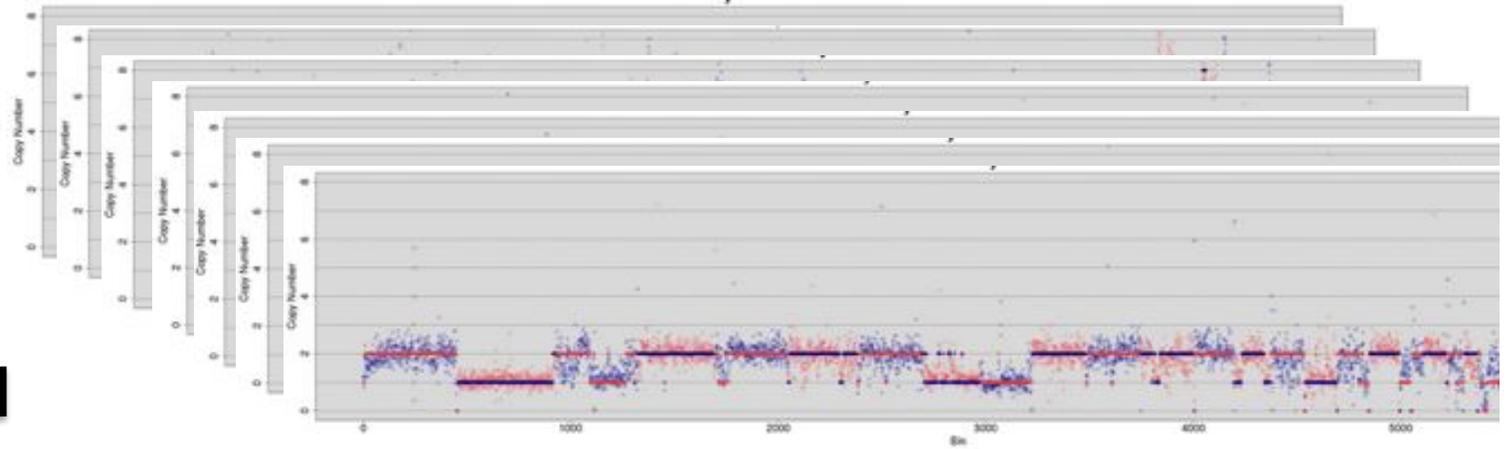
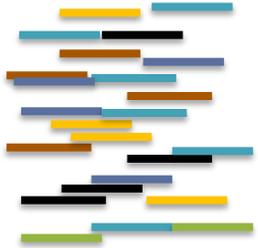
Structural Variation



Amplification

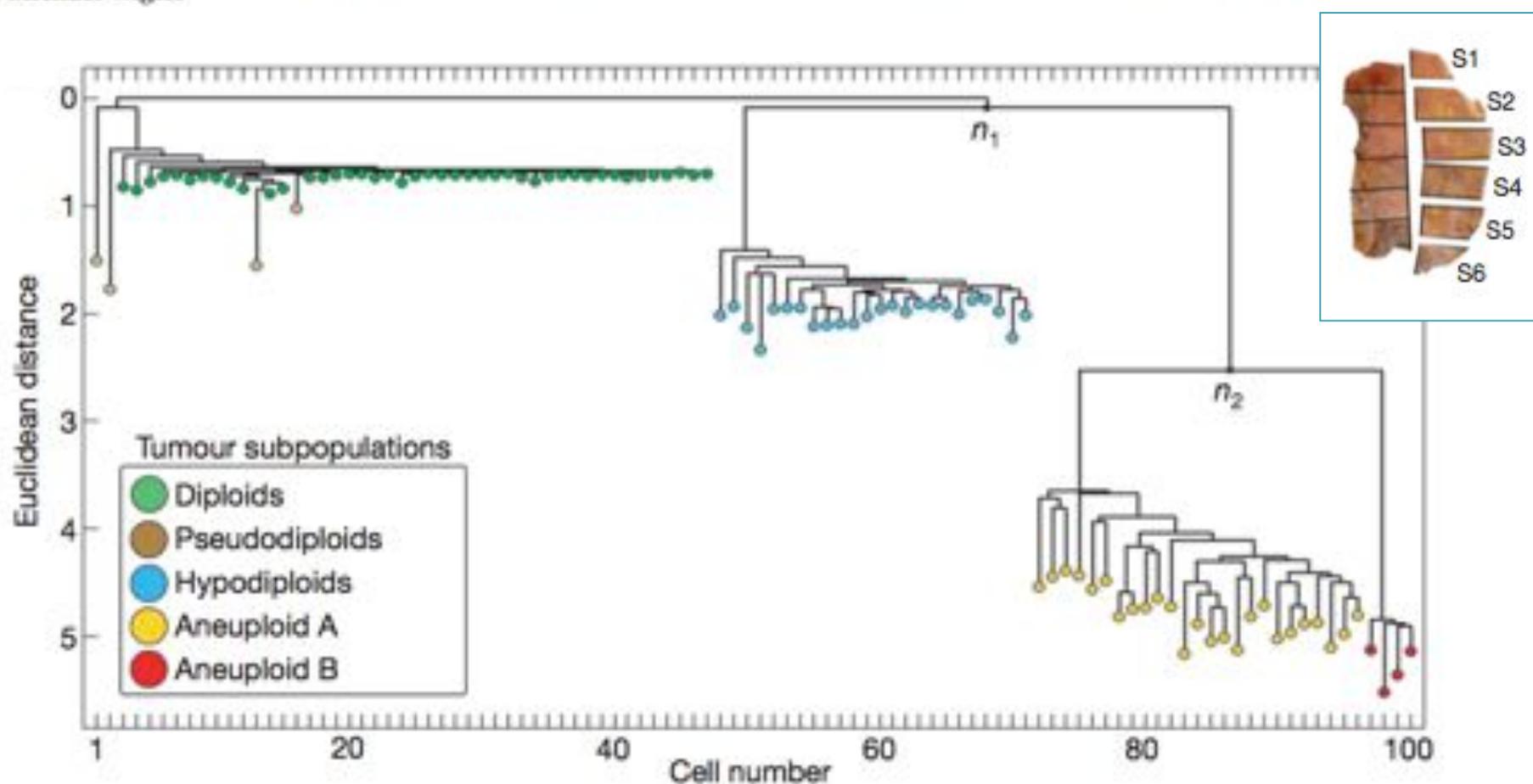


# CNV Analysis Overview



# Tumour evolution inferred by single-cell sequencing

Nicholas Navin<sup>1,2</sup>, Jude Kendall<sup>1</sup>, Jennifer Troge<sup>1</sup>, Peter Andrews<sup>1</sup>, Linda Rodgers<sup>1</sup>, Jeanne McIndoo<sup>1</sup>, Kerry Cook<sup>1</sup>, Asya Stepansky<sup>1</sup>, Dan Levy<sup>1</sup>, Diane Esposito<sup>1</sup>, Lakshmi Muthuswamy<sup>3</sup>, Alex Krasnitz<sup>1</sup>, W. Richard McCombie<sup>1</sup>, James Hicks<sup>1</sup> & Michael Wigler<sup>2</sup>



# Other Examples: CNV + RNA

PNAS

## Reproducible copy number variation patterns among single circulating tumor cells of lung cancer patients

Xiaohui Ni<sup>a,b,1</sup>, Minglei Zhuo<sup>c,1</sup>, Zhe Su<sup>a,1</sup>, Jianchun Duan<sup>c,1</sup>, Yan Gao<sup>a,1</sup>, Zhijie Wang<sup>c,1</sup>, Chenghang Zong<sup>b,1,2</sup>, Hua Bai<sup>c</sup>, Alec R. Chapman<sup>b,d</sup>, Jun Zhao<sup>c</sup>, Liya Xu<sup>a</sup>, Tongtong An<sup>c</sup>, Qi Ma<sup>a</sup>, Yuyan Wang<sup>c</sup>, Meina Wu<sup>c</sup>, Yu Sun<sup>e</sup>, Shuhang Wang<sup>c</sup>, Zhenxiang Li<sup>c</sup>, Xiaodan Yang<sup>c</sup>, Jun Yong<sup>b</sup>, Xiao-Dong Su<sup>a</sup>, Youyong Lu<sup>f</sup>, Fan Bai<sup>a,3</sup>, X. Sunney Xie<sup>a,b,3</sup>, and Jie Wang<sup>c,3</sup>



## Probing Meiotic Recombination and Aneuploidy of Single Sperm Cells by Whole-Genome Sequencing

Sijia Lu *et al.*  
*Science* **338**, 1627 (2012);  
DOI: 10.1126/science.1229112



## Mosaic Copy Number Variation in Human Neurons

Michael J. McConnell *et al.*  
*Science* **342**, 632 (2013);  
DOI: 10.1126/science.1243472



## Genome Analyses of Single Human Oocytes

Yu Hou,<sup>1,6</sup> Wei Fan,<sup>1,4,6</sup> Liying Yan,<sup>1,6</sup> Rong Li,<sup>1</sup> Ying Lian,<sup>1</sup> Jin Huang,<sup>1</sup> Jinsen Li,<sup>1</sup> Liya Xu,<sup>1</sup> Fuchou Tang,<sup>1,5,\*</sup> X. Sunney Xie,<sup>1,2,\*</sup> and Jie Qiao<sup>1,3,\*</sup>

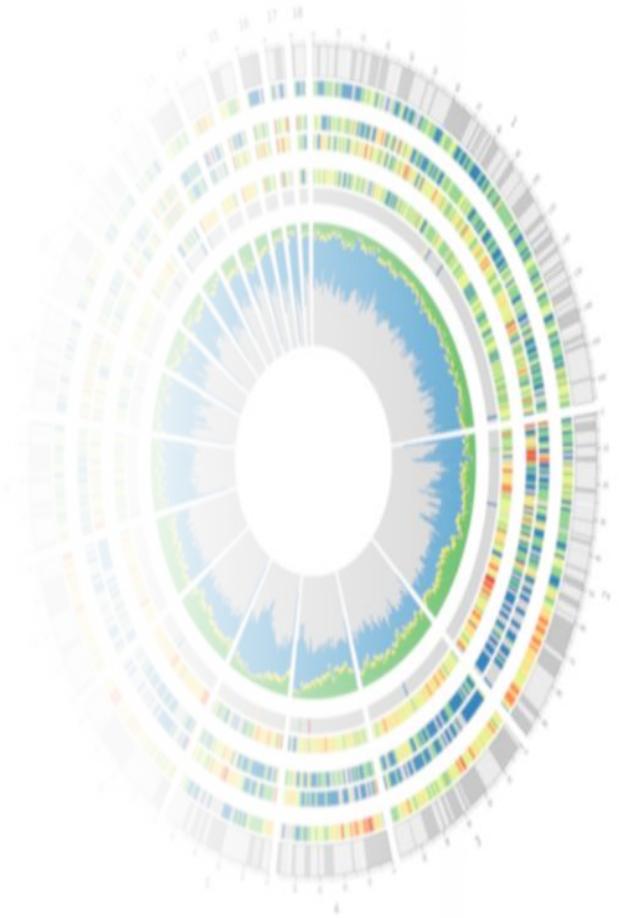
nature  
biotechnology

## The dynamics and regulators of cell fate decisions are revealed by pseudotemporal ordering of single cells

Cole Trapnell<sup>1,2,6</sup>, Davide Cacchiarelli<sup>1-3,6</sup>, Jonna Grimsby<sup>2</sup>, Prapti Pokharel<sup>2</sup>, Shuqiang Li<sup>4</sup>, Michael Morse<sup>1,2</sup>, Niall J Lennon<sup>2</sup>, Kenneth J Livak<sup>4</sup>, Tarjei S Mikkelsen<sup>1-3</sup> & John L Rinn<sup>1,2,5</sup>

# OMICS Summary

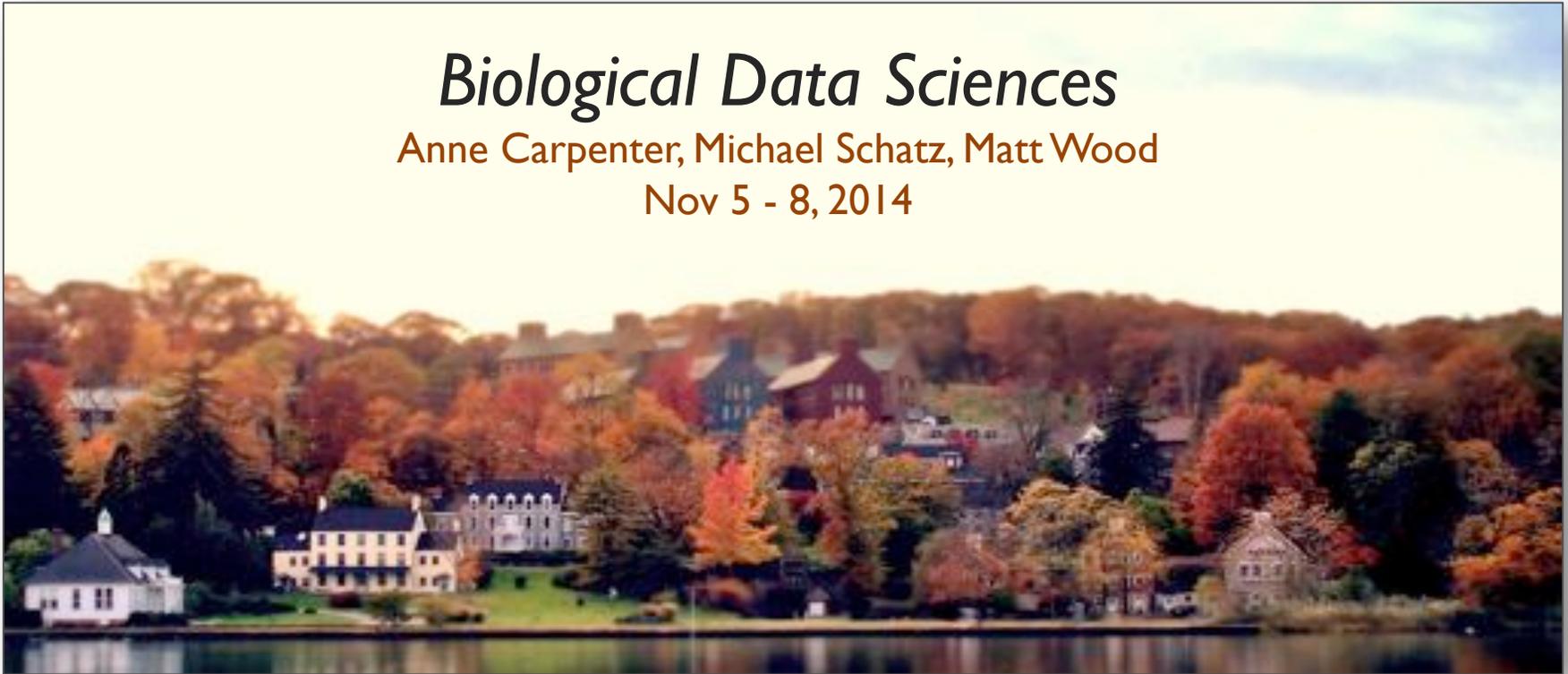
- DNA sequencing is extremely powerful and widespread to genotype large populations
  - The types of questions we ask have fundamentally changed over the last 10 years
  - Expect millions of human genomes over your PhD
- DNA sequencing is used for much more than sequencing DNA!
  - Flexible technology to observe the dynamics inside cells
  - Count the frequency of different molecules
  - See the “shadow” of chemical modification
  - See the “shadow” of molecules binding
- Coming up
  - Human Medical Genetics (Lyon)
  - Expression analysis (Gillis)
  - Genetics of modern and ancient humans (Schatz)
  - Group Discussion on ENCODE



# *Biological Data Sciences*

Anne Carpenter, Michael Schatz, Matt Wood

Nov 5 - 8, 2014



# Thank you!

<http://schatzlab.cshl.edu>

@mike\_schatz